

African Journal of Microbiology Research

Volume 11 Number 1 7 January, 2017

ISSN 1996-0808



*Academic
Journals*

ABOUT AJMR

The African Journal of Microbiology Research (AJMR) is published weekly (one volume per year) by Academic Journals.

The African Journal of Microbiology Research (AJMR) provides rapid publication (weekly) of articles in all areas of Microbiology such as: Environmental Microbiology, Clinical Microbiology, Immunology, Virology, Bacteriology, Phycology, Mycology and Parasitology, Protozoology, Microbial Ecology, Probiotics and Prebiotics, Molecular Microbiology, Biotechnology, Food Microbiology, Industrial Microbiology, Cell Physiology, Environmental Biotechnology, Genetics, Enzymology, Molecular and Cellular Biology, Plant Pathology, Entomology, Biomedical Sciences, Botany and Plant Sciences, Soil and Environmental Sciences, Zoology, Endocrinology, Toxicology. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles are peer-reviewed.

Contact Us

Editorial Office: ajmr@academicjournals.org

Help Desk: helpdesk@academicjournals.org

Website: <http://www.academicjournals.org/journal/AJMR>

Submit manuscript online <http://ms.academicjournals.me/>

Editors

Prof. Stefan Schmidt

*Applied and Environmental Microbiology
School of Biochemistry, Genetics and Microbiology
University of KwaZulu-Natal
Pietermaritzburg,
South Africa.*

Prof. Fukai Bao

*Department of Microbiology and Immunology
Kunming Medical University
Kunming,
China.*

Dr. Jianfeng Wu

*Dept. of Environmental Health Sciences
School of Public Health
University of Michigan
USA.*

Dr. Ahmet Yilmaz Coban

*OMU Medical School
Department of Medical Microbiology
Samsun,
Turkey.*

Dr. Seyed Davar Siadat

*Pasteur Institute of Iran
Pasteur Square, Pasteur Avenue
Tehran,
Iran.*

Dr. J. Stefan Rokem

*The Hebrew University of Jerusalem
Department of Microbiology and Molecular
Genetics
Jerusalem,
Israel.*

Prof. Long-Liu Lin

*National Chiayi University
Chiayi,
Taiwan.*

Dr. Thaddeus Ezeji

*Fermentation and Biotechnology Unit
Department of Animal Sciences
The Ohio State University
USA.*

Dr. Mamadou Gueye

*MIRCEN/Laboratoire commun de microbiologie
IRD-ISRA-UCAD
Dakar, Senegal.*

Dr. Caroline Mary Knox

*Department of Biochemistry, Microbiology and
Biotechnology
Rhodes University
Grahamstown,
South Africa.*

Dr. Hesham Elsayed Mostafa

*Genetic Engineering and Biotechnology Research
Institute (GEBRI)
Mubarak City For Scientific Research
Alexandria, Egypt.*

Dr. Wael Abbas El-Naggar

*Microbiology Department
Faculty of Pharmacy
Mansoura University
Mansoura, Egypt.*

Dr. Barakat S.M. Mahmoud

*Food Safety/Microbiology
Experimental Seafood Processing Laboratory
Costal Research and Extension Center
Mississippi State University
Pascagoula,
USA.*

Prof. Mohamed Mahrous Amer

*Faculty of Veterinary Medicine
Department of Poultry Diseases
Cairo university
Giza, Egypt.*

Editors

Dr. R. Balaji Raja

*Department of Biotechnology
School of Bioengineering
SRM University
Chennai,
India.*

Dr. Aly E Abo-Amer

*Division of Microbiology
Botany Department
Faculty of Science
Sohag University
Egypt.*

Editorial Board Members

Dr. Haoyu Mao

*Department of Molecular Genetics and Microbiology
College of Medicine
University of Florida
Florida, USA.*

Dr. Yongxu Sun

*Department of Medicinal Chemistry and
Biomacromolecules
Qiqihar Medical University
Heilongjiang
P.R. China.*

Dr. Ramesh Chand Kasana

*Institute of Himalayan Bioresource Technology
Palampur,
India.*

Dr. Pagano Marcela Claudia

*Department of Biology,
Federal University of Ceará - UFC
Brazil.*

Dr. Pongsak Rattanachaiakunsopon

*Department of Biological Science
Faculty of Science
Ubon Ratchathani University
Thailand.*

Dr. Gokul Shankar Sabesan

*Microbiology Unit, Faculty of Medicine
AIMST University
Kedah,
Malaysia.*

Editorial Board Members

Dr. Kamel Belhamef

*Faculty of Technology
University of Bejaia
Algeria.*

Dr. Sladjana Jevremovic

*Institute for Biological Research
Belgrade,
Serbia.*

Dr. Tamer Edirne

*Dept. of Family Medicine
Univ. of Pamukkale
Turkey.*

Dr. Mohd Fuat ABD Razak

*Institute for Medical Research
Malaysia.*

Dr. Minglei Wang

*University of Illinois at Urbana-Champaign
USA.*

Dr. Davide Pacifico

*Istituto di Virologia Vegetale – CNR
Italy.*

Prof. N. S. Alzoreky

*Food Science & Nutrition Department
College of Agricultural Sciences & Food
King Faisal University
Saudi Arabia.*

Dr. Chen Ding

*College of Material Science and Engineering
Hunan University
China.*

Dr. Sivakumar Swaminathan

*Department of Agronomy
College of Agriculture and Life Sciences
Iowa State University
USA.*

Dr. Alfredo J. Anceno

*School of Environment, Resources and Development (SERD)
Asian Institute of Technology
Thailand.*

Dr. Iqbal Ahmad

*Aligarh Muslim University
Aligrah,
India.*

Editorial Board Members

Dr. Juliane Elisa Welke

*UFRGS – Universidade Federal do Rio Grande do Sul
Brazil.*

Dr. Iheanyi Omezuruike Okonko

*Department of Virology
Faculty of Basic Medical Sciences
University of Ibadan
Ibadan,
Nigeria.*

Dr. Giuliana Noratto

*Texas A&M University
USA.*

Dr. Babak Mostafazadeh

*Shaheed Beheshty University of Medical Sciences
Iran.*

Dr. Mehdi Azami

*Parasitology & Mycology Department
Baghaeei Lab.
Isfahan,
Iran.*

Dr. Rafel Socias

*CITA de Aragón
Spain.*

Dr. Anderson de Souza Sant'Ana

*University of São Paulo
Brazil.*

Dr. Juliane Elisa Welke

*UFRGS – Universidade Federal do Rio Grande do Sul
Brazil.*

Dr. Paul Shapshak

*USF Health
Depts. Medicine and Psychiatry & Beh Med.
Div. Infect. Disease & Internat Med
USA.*

Dr. Jorge Reinheimer

*Universidad Nacional del Litoral (Santa Fe)
Argentina.*

Dr. Qin Liu

*East China University of Science and Technology
China.*

Dr. Samuel K Ameyaw

*Civista Medical Center
USA.*

Dr. Xiao-Qing Hu

*State Key Lab of Food Science and Technology
Jiangnan University
China.*

Prof. Branislava Kocic

*University of Nis
School of Medicine
Institute for Public Health
Nis,
Serbia.*

Prof. Kamal I. Mohamed

*State University of New York
Oswego,
USA.*

Dr. Adriano Cruz

*Faculty of Food Engineering-FEA
University of Campinas (UNICAMP)
Brazil.*

Dr. Mike Agenbag

*Municipal Health Services,
Joe Gqabi,
South Africa.*

Dr. D. V. L. Sarada

*Department of Biotechnology
SRM University
Chennai
India.*

Prof. Huaizhi Wang

*Institute of Hepatopancreatobiliary
Surgery of PLA Southwest Hospital
Third Military Medical University
Chongqing
China.*

Prof. A. O. Bakhiet

*College of Veterinary Medicine
Sudan University of Science and Technology
Sudan.*

Dr. Saba F. Hussain

*Community, Orthodontics and Paediatric Dentistry
Department
Faculty of Dentistry
Universiti Teknologi MARA
Selangor,
Malaysia.*

Editorial Board Members

Prof. Zohair I. F. Rahemo

*Department of Microbiology and Parasitology
Clinical Center of Serbia
Belgrade,
Serbia.*

Dr. Afework Kassu

*University of Gondar
Ethiopia.*

Dr. How-Yee Lai

*Taylor's University College
Malaysia.*

Dr. Nidheesh Dadheech

*MS. University of Baroda,
Vadodara,
India.*

Dr. Franco Mutinelli

*Istituto Zooprofilattico Sperimentale delle Venezie
Italy.*

Dr. Chanpen Chanchao

*Department of Biology,
Faculty of Science,
Chulalongkorn University
Thailand.*

Dr. Tsuyoshi Kasama

*Division of Rheumatology,
Showa University
Japan.*

Dr. Kuender D. Yang

*Chang Gung Memorial Hospital
Taiwan.*

Dr. Liane Raluca Stan

*University Politehnica of Bucharest
Department of Organic Chemistry
Romania.*

Dr. Mohammad Feizabadi

*Tehran University of Medical Sciences
Iran.*

Prof. Ahmed H Mitwalli

*Medical School
King Saud University
Riyadh,
Saudi Arabia.*

Dr. Mazyar Yazdani

*Department of Biology
University of Oslo
Blindern,
Norway.*

Dr. Babak Khalili Hadad

*Department of Biological Sciences
Islamic Azad University
Roudehen,
Iran.*

Dr. Ehsan Sari

*Department of Plant Pathology
Iranian Research Institute of Plant Protection
Tehran,
Iran.*

Dr. Snjezana Zidovec Lepej

*University Hospital for Infectious Diseases
Zagreb,
Croatia.*

Dr. Dilshad Ahmad

*King Saud University
Saudi Arabia.*

Dr. Adriano Gomes da Cruz

*University of Campinas (UNICAMP)
Brazil*

Dr. Hsin-Mei Ku

*Agronomy Dept.
NCHU
Taichung, Taiwan.*

Dr. Fereshteh Naderi

*Islamic Azad University
Iran.*

Dr. Adibe Maxwell Ogochukwu

*Department of Clinical Pharmacy and Pharmacy
Management,
University of Nigeria
Nsukka,
Nigeria.*

Dr. William M. Shafer

*Emory University School of Medicine
USA.*

Dr. Michelle Bull

*CSIRO Food and Nutritional Sciences
Australia.*

Editorial Board Members

Prof. Márcio Garcia Ribeiro

*School of Veterinary Medicine and Animal Science-UNESP,
Dept. Veterinary Hygiene and Public Health,
State of Sao Paulo
Brazil.*

Prof. Sheila Nathan

*National University of Malaysia (UKM)
Malaysia.*

Prof. Ebiamadon Andi Brisibe

*University of Calabar,
Calabar,
Nigeria.*

Dr. Julie Wang

*Burnet Institute
Australia.*

Dr. Jean-Marc Chobert

*INRA- BIA, FIPL
France.*

Dr. Zhilong Yang

*Laboratory of Viral Diseases
National Institute of Allergy and Infectious Diseases,
National Institutes of Health
USA.*

Dr. Dele Raheem

*University of Helsinki
Finland.*

Dr. Biljana Miljkovic-Selimovic

*School of Medicine,
University in Nis,
Serbia.*

Dr. Xinan Jiao

*Yangzhou University
China.*

Dr. Endang Sri Lestari, MD.

*Department of Clinical Microbiology,
Medical Faculty,
Diponegoro University/Dr. Kariadi Teaching Hospital,
Semarang
Indonesia.*

Dr. Hojin Shin

*Pusan National University Hospital
South Korea.*

Dr. Yi Wang

*Center for Vector Biology
Rutgers University
New Brunswick
USA.*

Prof. Natasha Potgieter

*University of Venda
South Africa.*

Dr. Sonia Arriaga

*Instituto Potosino de Investigación Científica y Tecnológica/
División de Ciencias Ambientales
Mexico.*

Dr. Armando Gonzalez-Sanchez

*Universidad Autonoma Metropolitana Cuajimalpa
Mexico.*

Dr. Pradeep Parihar

*Lovely Professional University
Punjab,
India.*

Dr. William H Roldán

*Department of Medical Microbiology
Faculty of Medicine
Peru.*

Dr. Kanzaki, L. I. B.

*Laboratory of Bioprospection
University of Brasilia
Brazil.*

Prof. Philippe Dorchies

*Nationale Veterinary School of Toulouse,
France.*

Dr. C. Ganesh Kumar

*Indian Institute of Chemical Technology,
Hyderabad
India.*

Dr. Zainab Z. Ismail

*Dept. of Environmental Engineering
University of Baghdad
Iraq.*

Dr. Ary Fernandes Junior

*Universidade Estadual Paulista (UNESP)
Brasil.*

Editorial Board Members

Dr. Fangyou Yu

*The first Affiliated Hospital of Wenzhou Medical College
China.*

Dr. Galba Maria de Campos Takaki

*Catholic University of Pernambuco
Brazil.*

Dr Kwabena Ofori-Kwakye

*Department of Pharmaceutics
Kwame Nkrumah University of Science & Technology
Kumasi,
Ghana.*

Prof. Liesel Brenda Gende

*Arthropods Laboratory,
School of Natural and Exact Sciences,
National University of Mar del Plata
Buenos Aires,
Argentina.*

Dr. Hare Krishna

*Central Institute for Arid Horticulture
Rajasthan,
India.*

Dr. Sabiha Yusuf Essack

*Department of Pharmaceutical Sciences
University of KwaZulu-Natal
South Africa.*

Dr. Anna Mensuali

*Life Science
Scuola Superiore Sant'Anna
Italy.*

Dr. Ghada Sameh Hafez Hassan

*Pharmaceutical Chemistry Department
Faculty of Pharmacy
Mansoura University
Egypt.*

Dr. Kátia Flávia Fernandes

*Department of Biochemistry and Molecular Biology
Universidade Federal de Goiás
Brasil.*

Dr. Abdel-Hady El-Gilany

*Department of Public Health & Community Medicine
Faculty of Medicine
Mansoura University
Egypt.*

Dr. Radhika Gopal

*Cell and Molecular Biology
The Scripps Research Institute
San Diego, CA
USA.*

Dr. Mutukumira Tony

*Institute of Food Nutrition and Human Health
Massey University
New Zealand.*

Dr. Habip Gedik

*Department of Infectious Diseases and Clinical
Microbiology
Ministry of Health Bakırköy Sadi Konuk Training and
Research Hospital
Istanbul,
Turkey.*

Dr. Annalisa Serio

*Faculty of Bioscience and Technology for Food
Agriculture and Environment
University of Teramo
Teramo,
Italy.*

ARTICLES

- Antimicrobial activity of *Phoma* sp. URM 7221: An endophyte from *Schinus terebinthifolius* Raddi (Anacardiaceae)** 1
Gustavo Bartolomeu Pedrosa Gomes da Silva, Karine Fagundes Silvino, Jadson Diogo Pereira Bezerra, Thaísa Gabriela Silva de Farias, Janete Magali de Araújo and Tânia Lúcia Montenegro Stamford
- A very high frequency of hepatitis B and C virus infections during an active screening campaign in Abidjan** 8
Kouassi-M'Bengue A., Ouattara Abdoulaye, Allah-Kouadio Emile, Sevede Daouda, Doumbia Moussa and Dosso M.
- Diversity and composition of methanotrophs in paddy soil as affected by different long-term fertilizer management from double-cropping paddy fields in Southern China** 16
Haiming Tang, Yilan Xu, Xiaoping Xiao, Jie Liu, Weiyang Li and Jimin Sun

Full Length Research Paper

Antimicrobial activity of *Phoma* sp. URM 7221: An endophyte from *Schinus terebinthifolius* Raddi (Anacardiaceae)

Gustavo Bartolomeu Pedrosa Gomes da Silva¹, Karine Fagundes Silvino¹, Jadson Diogo Pereira Bezerra², Thaísa Gabriela Silva de Farias³, Janete Magali de Araújo¹ and Tânia Lúcia Montenegro Stamford^{4*}

¹Departamento de Antibióticos, Universidade Federal de Pernambuco, Av. Professor Moraes Rego, 1235 - Cidade Universitária, Recife - PE, Brazil.

²Departamento de Micologia Prof. Chaves Batista, Universidade Federal de Pernambuco, Av. Professor Moraes Rego, 1235 - Cidade Universitária, Recife - PE, Brazil.

³Departamento de Nutrição, Universidade Federal de Pernambuco, Av. Professor Moraes Rego, 1235 - Cidade Universitária, Recife - PE, Brazil.

⁴Departamento de Nutrição, Universidade Federal de Pernambuco, Av. Professor Moraes Rego, 1235 - Cidade Universitária, Recife - PE, Brazil.

Received 5 October 2016, Accepted 2 December, 2016.

The discovery of new metabolites potentially bioactive against pathogenic microorganisms, mainly multidrug resistant, has aroused interest in endophytic fungi. The plant-associated microorganisms have been an important source for development of new compounds of biotechnological interest. This study aimed to investigate the antibacterial capacity of the endophytic fungus, *Phoma* sp. URM 7221 isolated from the medicinal plant *Schinus terebinthifolius* against human-pathogenic bacteria. An endophyte was isolated from *S. terebinthifolius* leaves. *Phoma herbarum* URM7221 was characterized morphologically and on the basis of ITS rDNA sequence. Primary antimicrobial activity was evaluated using the agar diffusion method and fermentation in liquid medium. Six different solvents were used to extract the active metabolites from fungal biomass and metabolic liquid. An antimicrobial activity test from the extract was carried out using a disk diffusion method with the endophytic extract containing the best antibacterial activity. Two tests were performed: Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC). Partially purified secondary metabolite extracts were analysed by thin layer chromatography (TLC). Liquid metabolically bioactive compounds extracted with petroleum ether revealed a MIC of 25 and 500 $\mu\text{g}\cdot\text{mL}^{-1}$ against *S. aureus* and MRSA, respectively. Ether and methanol extracts were assessed by chemical analyses and contained phenolic compounds, triterpenes, steroids, reducing sugars, mono- and sesquiterpenes. The thin layer chromatography assay showed the activity of different antimicrobial compounds produced by *Phoma* sp. URM7221. This endophyte (URM7221) could be efficiently used for production of bioactive metabolites against pathogenic microorganisms, with significant biotechnological potential.

Key words: Bioactive compounds, endophytic fungi, pathogenic bacteria, multidrug-resistance, antibacterial agents.

INTRODUCTION

The overuse of antibiotics in medical practice has contributed to the increase in antibiotic-resistant microorganisms (Paes et al., 2014). Infections caused by Gram-positive bacteria with antibiotic resistance are one of the main reasons for the high mortality and morbidity rates reported for patients, resulting in costly treatment (Woodford and Livermore, 2009). *Staphylococcus aureus* is one of the most common pathogen in a diverse number of infectious diseases, including simple skin infections and invasive diseases that lead to bacteremia and sepsis (Kim et al., 2012; Rocha et al., 2015). This bacterium requires attention due to emergence of resistant strains to the primary antibiotics used in their treatment, such as methicillin. Methicillin-resistant *S. aureus* (MRSA) is a major cause of hospital morbidity and mortality (Siqueira et al., 2011). Similar to *S. aureus*, the genus *Enterococcus* includes some of the most important opportunistic pathogens. Their variable genome has influence on their adaptation to different environments (Arias and Murray, 2012).

The race to reduce the impact of infections caused by multidrug resistant bacteria, has reinforced the need to discover new antimicrobial substances. Studies have researched new compounds in plants and microorganisms that live inside vegetal tissues, such as endophytes. Endophytes are microorganisms that live in association with plant tissue (Kusari et al., 2012) and many species are known to produce bioactive secondary metabolites with medicinal potential (Kusari et al., 2012). This suggests alternative paths to discovery of new bioactive substances, leading to development of new drugs derived from compounds produced by endophytes species (Kumar et al., 2011).

Several studies have demonstrated that endophytic fungi can produce numerous compounds with biological activities of interest, such as antitumor (Jin-long et al., 2011; Chandra, 2012), antimicrobial (Siqueira et al., 2011; Tayung et al., 2012; Bagchi and Banerjee, 2013; Pinheiro et al., 2013; Orlandelli et al., 2015), enzymes (Chandra, 2012), plant growth hormones (Hwang et al., 2011), leishmanicidal (Santiago et al., 2012), cytotoxic compounds (Li et al., 2011) and compounds with industrial and pharmaceutical potential (Meng et al., 2011; Wang and Dai, 2011), reinforcing the broad biotechnological potential of these microorganisms.

Schinus terebinthifolius Raddi belongs to Anacardiaceae family and is largely found in the coastal region of Brazil (Carvalho et al., 2013). This plant has

used in traditional medicine as antipyretic, analgesic and in the treatment of urogenital diseases (Carvalho et al., 2013). Several studies have shown that *S. terebinthifolius* produce pharmacologically important substances, such as anticancer (Bendaoud et al., 2010; Matsuo et al., 2011), antimicrobial (Silva et al., 2010; Pereira et al., 2011), antioxidant (Bendaoud et al., 2010), healing purposes (Estevão et al., 2013) and antiproliferative (Queires et al., 2013).

Considering the impact on health human and the necessity of more studies verifying the biotechnological properties of endophytic fungi, the present study aims to 1) evaluate the antibacterial potential of endophytic fungi against Gram-positive pathogenic bacteria and 2) identify the class of bioactive metabolites produced by *Phoma* sp. (URM 7221) by thin layer chromatography (TLC).

MATERIALS AND METHODS

Endophytic fungi: Isolation and identification

S. terebinthifolius Raddi leaves were collected from different plants on the campus of Federal University of Pernambuco, Recife city, Pernambuco state, Brazil (08°03'07"S 34°56'59"O). For endophytic isolation, healthy leaves were selected as described by McInroy et al. (1995). The leaves were cut into 180 fragments of approximately 1 cm² and were cultured for up to 20 days at 30°C on Sabouraud agar and potato dextrose agar (PDA) media containing chloramphenicol (100 mg/L).

For morphological identification of endophytic fungi, the fungi were cultured on Synthetic Nutrient-poor Agar (SNA), malt extract agar (MEA) and potato dextrose agar (PDA) media for 10 days in the dark at 28°C and for 10 days under continuous UVA light. After this period of cultivation macro and microscopic characteristics of colonies were observed according to Boerema et al. (2004). For molecular analysis, 7-day old culture on PDA was used for DNA extraction with an UltraClean™ Microbial Isolation Kit (Mo Bio Laboratories, Carlsbad, CA, USA) according to the manufacturer's instructions.

Polymerase chain reaction for the part of ITS region (first and second internal transcribed spacer regions and intervening 5.8S nrDNA) amplification was performed with 50 µL volume, using Taq® DNA polymerase 1X buffer, 1.5 mM MgCl₂, 0.4 µM ITS1 and ITS4 primers (White et al., 1990), 0.2 mM dNTPs, 0.2 Taq® DNA polymerase and 25ng DNA. Amplification was done in a thermal cycler with the following conditions: 5 min at 95°C (1 cycle), 30 s at 95°C (30 cycles), 1 min at 62°C (annealing), 2 min at 72 °C (extension) and 5 min at 72°C (final extension). Amplification products were separated by electrophoresis on a 1% agarose gel, colored with GelRed® and visualized with UV light using molecular weight marker 1 kb plus (Fermentas®). PureLink PCR Purification Kit (Invitrogen®) was used according to manufacturer's recommendation. Amplification products were sequenced and electropherograms were edited by Staden Package software. The consensus sequence was blasted against sequences on GenBank

*Corresponding author. E-mail: flmstamford@yahoo.com.br. Tel: +55 81 2126-8470. Fax: +55 81 2126-8474.

using BLASTn. Sequences with the highest similarity percentage (99-100%) in comparison with the studied sequence were analysed.

Antimicrobial activity: Solid medium

Forty endophytic fungi isolated with distinct morphological characteristics were selected for antimicrobial activity. Four microorganisms were tested using an agar block test as previously described by Ichikawa et al. (1971). The four bacteria species used for the test were: *S. aureus* Rosenbach (UFPEDA02), MRSA (UFPEDA663), *Enterococcus faecalis* (Andrewes & Horder) Schleifer & Kilpper-Bälz (UFPEDA138), and *Bacillus subtilis* (Ehrenberg) Cohn (UFPEDA86). The inhibition zones obtained were compared with control information from the table of the Clinical and Laboratory Standards Institute (CLSI, 2013).

Fermentation in liquid medium

The endophytic fungus (URM 7221) that demonstrated the best antimicrobial activity using solid medium was selected to the test in liquid medium for extraction of active metabolites. After growth, fragments of 10 mm diameter of the endophytes were transferred to Erlenmeyer flasks (250 mL) with 50 mL of the liquid medium MPE (Hamada et al., 1974), Czapeck and M1 (20 g/L glucose, 200 g/L peptone) incubated for 2 days, at 30°C at 120 rpm. After this period, 10 mL of each pre-inoculum was transferred to flasks (500 mL) containing 90 mL of the same liquid medium and incubated in the same conditions. Every 24 h (during 120 h), 1 mL of fermentation broth was removed from the flask and centrifuged at 10.000 rpm for 3 min. From the supernatant fluid, 50 µL was used to perform the disk diffusion antimicrobial test (Kirby et al., 1966).

Endophytic extracts

After fermentation of endophytic fungus (URM 7221) for 48 h in MPE medium, the fungal biomass was separated from the metabolic liquid using Whatman filter paper 4 and centrifuged at 10.000 rpm for 3 min. Ethanol, methanol and acetone (1:10 g/mL) at pH 2.0, 7.0 and 9.0, respectively, were used to extract the active metabolites from the biomass. To extract the active substances present in the metabolic liquid petroleum ether, ethyl acetate and chloroform (2:1) were used. The pH of the metabolic liquid was adjusted to 2.0, 7.0 and 9.0, respectively. After 1 h of shaking, the samples were centrifuged at 10,000 rpm for 3 min and the solvent containing the extract was evaporated using a rotary evaporator at 55°C (Lyra et al., 1964).

Antimicrobial activity: Endophytic extract

The antimicrobial activity test of the endophytic extract was performed using a disk diffusion method, with sterile paper discs impregnated with 50 µL of each extract. The pH of the endophytic extract was adjusted to 7.0, to avoid interferences. The diameter of the inhibition zones around the discs was measured and was assessed using the same microorganisms tests as described in "Antimicrobial activity: solid medium" section. Chloramphenicol 1 mg/mL was used as a positive control, methanol was applied as solvent control, and the depleted liquid and water were used as negatives controls (Lyra et al., 1964).

Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC)

Two tests- MIC and MBC were used to assay the endophytic extract with the best antibacterial activity. A broth microdilution assay using 96-well microplate, performed according to the Clinical and Laboratory Standards Institute (CLSI, 2013) was used. All determinations were executed in triplicate.

Plant tissue extraction

For extraction of compounds with no antimicrobial activity, leaves of *S. terebinthifolius* (1 g) were immersed in 10 mL of hexane for two hours. The leaves were then immersed in 10 mL of ethyl acetate for two hours to extract bioactive compounds. Solvent was evaporated using a rotary evaporator at 55°C (Ceruks et al., 2007).

Secondary metabolic analyses

For partial purification of secondary metabolites, 10 µL aliquots of biomass extract (methanol) and metabolic fluid extract (ether) of *Phoma* sp. (URM 7221) were analysed by thin layer chromatography (TLC) on silica gel F254 (Merck®). Several mobile phases, such as ethyl acetate : acetic acid : formic acid : water (100:11:11:26, v/v), toluene : ethyl acetate (80:20; 97:3, v/v), acetone : n-butanol : phosphate buffer pH 5.0 (5:4:1, v/v), and specific visualization reagents were used (Robertson et al., 1956; Metz, 1961; Sharma and Darwra, 1991; Wagner and Bladt, 1996; Harbone, 1998). The chromatograms were run in saturated chambers.

The bioactive extracts obtained from *Phoma* sp. (URM 7221) were compared with compounds present in crude ethyl acetate extract of leaves by TLC using an agar overlay method (Rodrigues et al., 2009). Ethyl acetate: methanol (9:1) solution was used as mobile phase. An aqueous solution of 2,3,5-triphenyltetrazolium chloride (20 mg/mL) (Rodrigues et al., 2009) was used for visualization. Biological activity was determined by the formation of white and well defined inhibition zones against a red-purple background. Retention factor (Rf) values were measured and compared (Homans and Fuchs, 1970). All tests were performed in triplicate.

RESULTS

Endophyte isolation, identification and antimicrobial activity

Using culture media for isolation of endophytes, 220 endophytic fungi were isolated from leaves of *S. terebinthifolius*. Among these, 137 endophytes (64.62%) were obtained using Sabouraud agar medium and 75 (35.37%) using PDA medium.

The endophyte that showed the best antibacterial activity was identified. Morphological analysis was performed after 20 days culture on SNA, MEA and PDA media. The fungus grew on three different media with similar macroscopic characteristics (velvety-powdered texture, colony color ranging between light brown and dark pink, reverse light brown to reddish brown). Pycnidia

Table 1. Antibacterial activity of endophytic fungi isolated from leaves of *S. terebinthifolius* against human pathogenic bacteria.

Test microorganism	Inhibition zones (mm)						
	Endophytic fungi						
	F-10	F-61 (URM 7221)	F-100	F-105	F-169	F-178	F-188
<i>Bacillus subtilis</i> UFPEDA86	20 ± 0.0	35 ± 0.0	16,4 ± 0,94	16 ± 0.8	-	-	>10 ± 0.0
<i>Enterococcus faecalis</i> UFPEDA138	17 ± 0.0	23.7 ± 0.47	16.7 ± 0.47	>10 ± 0.0	-	>10 ± 0.0	-
<i>Staphylococcus aureus</i> UFPEDA02	-	35 ± 0.0	-	>10 ± 0.0	19 ± 0.0	>10 ± 0.0	>10 ± 0.0
<i>Staphylococcus aureus</i> (MRSA) UFPEDA663	16 ± 0.86	22,4 ± 0,47	-	-	-	-	>10 ± 0.0

F: Endophytic isolates; (MRSA): Methicillin-resistant *Staphylococcus aureus* resistant; (-): no activity.

Table 2. Susceptibility profile of test microorganisms to endophyte *Phoma* sp. (URM 7221) grown in MPE or M1 liquid fermentation.

Test Microorganisms	Inhibition zones (mm) over incubation time									
	24 h		48 h		72 h		96 h		120 h	
	MPE	M1	MPE	M1	MPE	M1	MPE	M1	MPE	M1
<i>B. subtilis</i>	14 ± 0.0	-	16.3 ± 0.94	17 ± 0.81	25 ± 0.94	12 ± 0.81	22.3 ± 0.47	15 ± 0.0	22.3 ± 0.47	15.6 ± 0.47
<i>S. aureus</i>	15 ± 0.0	-	31.3 ± 0.47	21 ± 0.47	29.3 ± 0.47	11 ± 0.81	15.3 ± 0.47	-	-	-
<i>S. aureus</i> MRSA	15 ± 0.0	-	26.4 ± 0.81	-	15 ± 0.81	-	-	-	-	-

-: No activity.

(91-111 × 58-203 μm) and conidia (4-5.5 × 2-2.5 μm) were formed only on SNA medium. Using ITS rDNA sequence to a megablast search of the NCBI GenBank nucleotide database, our sequence (KP966098) has high identity (98 to 100%) to sequences deposited as *Phoma* sp. and *P. herbarum* (HQ630963, Shrestha et al., 2011; KJ188712, Luo et al., 2014), among others. The morphological characters described above match the *P. herbarum* description by Boerema (2004). However, because the new publications (Chen et al., 2015) using multigene phylogenetic analyses on the taxonomy of this genus, the authors decided to identify the isolate only as *Phoma* sp. (URM 7221).

Forty endophytes were tested in the primary assay, and only seven (17.5%) expressed antimicrobial activity. The endophyte identified as *Phoma* sp. (URM 7221) showed the greatest activity against *B. subtilis* (inhibition zone of 35 mm), *E. faecalis* (23.7 mm), *S. aureus* (35 mm) and MRSA (22.4 mm) (Table 1).

Fermentation and endophytic extracts antimicrobial activity

Fermentation tests revealed that MPE medium was the most efficient and only Czapeck medium did not stimulate the production of active metabolites against the four bacteria tested. *E. faecalis* (UFPEDA138) was resistant

to all endophytic extracts tested. The most efficient extract (using MPE medium) gave the best production of the bioactive principle and the largest inhibition zones against *S. aureus* (31 ± 0.47 3 mm after 48 h), MRSA (26.4 ± 0.81 mm, 48 h) and *B. subtilis* (25 ± 0.94 mm, 72 h). These results indicate that the production of bioactive metabolites with antimicrobial activity changes during incubation time (Table 2).

Antibacterial activity of biomass and metabolic liquid extracts obtained from *Phoma* sp. (URM 7221) and from leaves of *S. terebinthifolius* was evaluated (Table 3). The endophyte was subjected to liquid fermentation in MPE medium (pH 7.0) at 48 h. Petroleum ether (pH 7.0) was the most efficient solvent because it extracted bioactive metabolites active against *S. aureus* and MRSA, with inhibition zones reaching up to 23 mm. Ethyl acetate was able to partially extract bioactive compounds against the test microorganisms with inhibition zones between 10-18.6 mm. MIC and MBC values of petroleum ether extract were, respectively, 125 and 250 mg L⁻¹ against *S. aureus*, and 500 and 1000 mg L⁻¹ against MRSA.

Ethanol and methanol were used to extract active metabolites from fungal biomass and were effective against *S. aureus* and MRSA. Methanol (pH 2.0 and 7.0) showed inhibition zones ranging from 12 to 15 mm, with MIC and MBC of 250 and 500 mg L⁻¹, for *S. aureus* and 1000 mg L⁻¹ and >1000 mg L⁻¹ against MRSA. Acetone extracts were only active against *S. aureus*.

Table 3. Antimicrobial activity of crude extracts of *Phoma* sp. (URM 7221) obtained from liquid fermentation. MLE: Metabolic liquid exhausted; EXT: extract.

	Solvents		Inhibition zones (mm)					
			<i>S. aureus</i> (UFPEDA 02)			MRSA (UFPEDA 663)		
			pH2	pH7	pH9	pH2	pH7	pH9
Biomass	Acetone	EXT	21.6 ± 0.47	25 ± 0.0	22 ± 0.81	15 ± 0.0	17 ± 0.0	-
	Ethanol	EXT	23 ± 0.0	24 ± 0.0	12 ± 0.0	13.6 ± 0.47	11 ± 0.0	-
	Methanol	EXT	21.3 ± 0.94	21.6 ± 1.2	21 ± 0.0	17 ± 0.0	17 ± 0.0	17 ± 0.81
Metabolic liquid	Ethyl acetate	EXT	27.6 ± 0.94	28 ± 0.81	27 ± 0.81	21.6 ± 0.47	20 ± 0.0	20.6 ± 0.47
		MLE	-	-	-	-	-	-
	Chloroform	EXT	26.6 ± 0.47	25.6 ± 0.47	25.6 ± 0.47	19 ± 0.0	19 ± 0.0	19 ± 0.0
		MLE	-	-	-	11 ± 0.0	-	-
	Petroleum ether	EXT	-	11.6 ± 0.47	-	-	12 ± 0.0	-
		MLE	24 ± 0.0	24 ± 0.0	23.6 ± 0.47	19 ± 0.0	17.3 ± 0.47	17 ± 0.0

Chemical comparison of active compounds

Analysis of metabolic liquid by TLC suggested the presence of phenolic compounds, triterpenes, steroids, mono and sesquiterpenes. Analysis of fungal biomass showed reducing sugars and phenolic compounds. Using bioautographic assay, both extracts showed antimicrobial activity against *S. aureus* and MRSA. Petroleum ether extract showed Rf values of 0.65 and 0.6 for *S. aureus* and MRSA, respectively. Similar Rf values were observed for methanol extracts (0.125 and 0.15).

DISCUSSION

Studies of medicinal plants have received wide attention in recent years due to the large diversity of endophytic microorganisms that inhabit plant tissues (Siqueira et al., 2011). New species, useful for biotechnology applications, have been reported to produce extracellular active metabolites with pharmacological interest (Bagchi and Banerjee, 2013; Pinheiro et al., 2013). These microorganisms can also protect plants against phytopathogens, temperature, drought and world weather changes (Gundel et al., 2010). This research demonstrates the diversity of endophytic fungi living in leaves of *S. terebinthifolius* and the capacity of these endophytes to produce molecules with biotechnological importance. In this study, a unique endophytic species identified as *Phoma* sp. (URM 7221) produced more efficiently bioactive compounds against four Gram-bacteria species.

The genus *Phoma* comprises several species and varieties that are recognized as producers of antimicrobial compounds (Bezerra et al., 2015). Researchers have

reported that *Phoma* species can be isolated from many plants, such as *Laguncularia racemosa* (Costa et al., 2012), *Amaranthus cruentus* (Pusz et al., 2015), *Taraxacum mongolicum* (Zhang et al., 2013a) and *Mitrajyna javanica* (Pharamat et al., 2013). An important taxonomic paper from CBS-KNAW (The Netherlands) highlights the importance of a polyphasic approach to characterise *Phoma* species and similar genera, and it presented an overview of the phytopathological importance of this genus (Aveskamp, 2010). This antimicrobial potential can be associated with production of phenolic secondary metabolites and steroids (Hwang et al., 2011). Similar to our results, these compounds have been identified by chemical prospecting, which might be related to antimicrobial activity presented by *Phoma* sp. (URM 7221).

According to Strobel et al. (2011), species of the genus *Phoma* can produce organic volatile compounds, of which sesquiterpenes are the most prominent. Hamayun et al. (2009) described that *P. herbarum* produced several metabolic compounds, because it is adaptable to many environmental conditions. These compounds can play a role in immunomodulation (Zhang et al., 2013b; Shen et al., 2014) and antitumor activity (Fang et al., 2011; Pharamat et al., 2013), which reinforces the importance of new surveys involving this fungal genus. Studies in different countries have tested *Phoma* spp. antimicrobial activity against Gram-positive bacteria. Some authors studying this genus reported similar results. For example, Kumar et al. (2010) tested culture filtrate with the capacity to inhibit *S. aureus* (11 to 19 mm). In this study, primary assays proved antimicrobial activity of *Phoma* sp. (URM 7221), efficiently inhibiting *S. aureus* (11 to 35 mm) and MRSA (15 to 26.4 mm). In addition to the initial tests that showed activity against *B.*

subtilis (12 - 35 mm) and *E. faecalis* (23.7 ± 0.47 mm), other experiments demonstrated that *Phoma* sp. (URM 7221) extracts were not able to suppress the growth of these bacteria. Shukla et al. (2014) also demonstrated antimicrobial potential of this endophyte against *S. aureus* (18.3 ± 0.10 mm). Similar results were obtained by Zhang et al. (2013a) in a study of fungi associated with *Taraxacum mongolicum*, and the authors also found positive results against *S. aureus* (40 mm). Shen et al. (2012) has isolated endophytes from branches of *Phyllostachys edulis* and tested *P. herbarum* extracts against bacteria; it was not able to inhibit *S. aureus* and *B. subtilis*. The same fact occurred according to Pharamat et al. (2013) when evaluating fungal endophytes species obtained from *Mitrajyna javanica*.

Conclusions

Study of endophytic fungi from leaves of *S. terebinthifolius* revealed that different plants are capable of harboring endophytes, such as *Phoma* sp. (URM 7221) that demonstrated the most efficient antibacterial activity among other endophytes.

ACKNOWLEDGEMENTS

The authors acknowledge Fundação de Amparo à Ciência e Tecnologia do Estado de Pernambuco (FACEPE) for financial support.

Conflict of interest

The authors declare that there is no conflict of interest.

REFERENCES

- Arias CA, Murray BE. (2012). The rise of the *Enterococcus*: beyond vancomycin resistance. *Nat. Rev. Microbiol.* 10:266-78.
- Aveskamp MM, Gruiter J, Woudenberg JHC (2010). Highlights of the Didymellaceae: A polyphasic approach to characterise *Phoma* and related pleosporalean genera. *Stud. Mycol.* 65:1-60.
- Bagchi B, Banerjee D (2013). Diversity of fungal endophytes in *Bauhinia vahlii* (a lianas) from different regions of Paschim Medinipur district of West Bengal. *Int. J. Sci. Environ. Technol.* 2:748-756.
- Bendaoud H, Romdhane M, Souchard JP (2010). Chemical Composition and Anticancer and Antioxidant Activities of *Schinus Molle* L. and *Schinus Terebinthifolius* Raddi Berries Essential Oils. *J. Food Sci.* 75:466-72.
- Bezerra JDP, Nascimento CCF, Barbosa RN (2015). Endophytic fungi from medicinal plant *Bauhinia forficata*: Diversity and biotechnological potential. *Braz. J. Microbiol.* 46:49-57.
- Bezerra JDP, Santos MGS, Barbosa RN (2013). Fungal endophytes from cactus *Cereus jamacaru* in Brazilian tropical dry forest: A first study. *Symbiosis*, 60:53-63.
- Boerema GH, De Gruyter J, Noordeloos ME (2004). *Phoma* identification manual: differentiation of specific and infra-specific taxa in culture. Wallingford, United Kingdom: CAB International.
- Carvalho MG, Melo AGN, Aragão CFS (2013). *Schinus terebinthifolius* Raddi: chemical composition, biological properties and toxicity. *Rev. Bras. Plantas Med.* 15:158-69.
- Ceruks M, Romoff P, Fávero AO . (2007). Constituintes fenólicos polares de *Schinus terebinthifolius* Raddi (Anacardiaceae). *Química Nova*, 30:507-599.
- Chandra S (2012). Endophytic fungi: novel sources of anticancer lead molecules. *Appl. Microbiol. Biot.* 95:47-59.
- Chen Q, Zhang G, Jiang J, Cai L, Crous PW. (2015). Resolving the *Phoma* enigma. *Stud. Mycol.* 82:137-217.
- Clinical and Laboratory Standards Institute (CLSI) (2013). Performance Standards For Antimicrobial Susceptibility Testing, Twenty-Third Informational Supplement, M100-S21. *Clin. Lab Stand Inst.* 32:1-184.
- Costa IPMW, Maia LC, Cavalcanti MA (2012). Diversity of leaf endophytic fungi in mangrove plants of Northeast Brazil. *Braz. J. Microbiol.* 43:1165-1173.
- Estevão LR, Mendonça FD, Baratella-Evêncio L, Simões RS, Barros ME, Arantes RM, Rachid MA, Evêncio-Neto J (2013). Effects of aroeira (*Schinus terebinthifolius* Raddi) oil on cutaneous wound healing in rats. *Acta Cir. Bras.* 3:202-209.
- Fang MJ, Fang H, Li WJ (2011). A new diphenyl ether from *Phoma* sp. strain, SHZK2. *Nat. Prod. Res.* 26:1224-8.
- Gundel PE, Martínez GMA, Batista WB (2010). Dynamics of Neotyphodium endophyte infection in ageing seed pools: incidence of differential viability loss of endophyte, infected seed and non-infected seed. *Annals Appl. Biol.* 156:199-209.
- Hamada M, Kondo S, Yokoyama T (1974). Minosaminomycin, a new antibiotic containing myo-inosamine. *J. Antibiot.* 27:81-83.
- Hamayun M, Khan SA, Khan AL et al. (2009). *Phoma* sp. as a new gibberellin-producing and plant growth-promoting fungus. *J. Microbiol. Biotechnol.* 19:1244-1249.
- Harbone JB. (1998). *Phytochemical methods: a guide to modern techniques of plant analysis*. New York, United States: Chapman and Hall.
- Homans AL, Fuchs A. (1970.) Direct bioautography in thin-layer chromatograms as a method for detecting fungitoxic substances. *J. Chromatogr.* 51:327-329.
- Hwang JS, You YH, Bae JJ (2011). Effects of endophytic fungal secondary metabolites on the growth and physiological response of *Carex kobomugi* Ohwi. *J. Coastal Res.* 27:544-548.
- Ichikawa T, Date M, Ishikura T (1971). An improvement of kasugamycin – Producing Strain by the Agar Piece Method and Prototroph Method. *Folia Microb.* 16:218-224.
- Jin-long C, Shun-Xing G, Pei-gen X (2011). Antitumor and antimicrobial activities of endophytic fungi from medicinal parts of *Aquilaria sinensis*. *J. Zhejiang Univ. Sci. B.* 12:385-392.
- Kim HK, Thammavongsa V, Schneewind O (2012). Recurrent infections and immune evasion strategies of *Staphylococcus aureus*. *Curr. Opin. Microbiol.* 15(1):92-99.
- Kirby WMMAW, Bauer J C, Sherris MT. (1966). Antibiotic susceptibility testing by a standardized single disc method. *Am. J. Clin. Pathol.* 45:493-499.
- Kumar CG, Mongolla P, Joseph J (2010). Antimicrobial activity from the extracts of fungal isolates of soil and dung samples from Kaziranga National Park, Assam, India. *J. Med. Mycol.* 20:283-289.
- Kumar U, Singh A, Sivakumar T (2011). Isolation and screening of endophytic actinomycetes from different parts of *Embllica officinalis*. *Pharm. Sci.* 2:423-34.
- Kusari S, Hertweck C, Spiteller M. (2012). Chemical ecology of endophytic fungi: Origins of secondary metabolites. *Chem. Biol.* 19:792-798.
- Li H, Huang H, Shao C (2011). Cytotoxic Norsesquiterpene Peroxides from the Endophytic Fungus *Talaromyces flavus* Isolated from the Mangrove Plant *Sonneratia apetala*. *J. Nat. Prod.* 74:1230-1235.
- Luo J, Walsh E, Naik A (2014). Temperate Pine Barrens and Tropical Rain Forests Are Both Rich in Undescribed Fungi. *PLoS ONE* 9:e103753.
- Lyra FDA, Gonçalves LO, Coelho JSB (1964). Ciclamina e

- cicladina, dois novos antibióticos produzidos pelo *Streptomyces capoamus* nov. sp. Anais Acad. Bras. Ciênc. 36:323-334.
- Matsuo AL, Figueiredo CR, Arruda DC (2011). α -Pinene isolated from *Schinus terebinthifolius* Raddi (*Anacardiaceae*) induces apoptosis and confers antimetastatic protection in a melanoma model. Biochem. Biophys. Res. Commun. 411:449-454.
- McInroy JA, Kloepper JWA. (1995). Survey of indigenous bacterial endophytes from cotton and sweet corn. Plant soil 173:337-342.
- Meng L, Sun P, Tang H (2011). Endophytic fungus *Penicillium chrysogenum*, a new source of hypocrellins. Biochem. Syst. Ecol. 39: 163-165.
- Metz H (1961). Thin-layer chromatography for rapid assays of enzymic steroid transformations. *Naturwissenschaften*, 48:569-570.
- Orlandelli RC, Almeida TT, Alberto RN (2015). Antifungal and proteolytic activities of endophytic fungi isolated from *Piper hispidum* Sw. Braz. J. Microbiol. 46:359-366.
- Paes ARM, Câmara JT, Santos DAS (2014). Epidemiological study of cross infection in Intensive Care Unit. Rev. Enf. Piauí 3:10-17.
- Pereira EM, Gomes RT, Freire NR et al. (2011). In vitro antimicrobial activity of Brazilian medicinal plant extracts against pathogenic microorganisms of interest to dentistry. *Planta Med.* 77:401-404.
- Pharamat T, Palaga T, Piapukiew J et al. (2013). Antimicrobial and anticancer activities of endophytic fungi from *Mitrajyna javanica* Koord and Val. Afr. J. Microbiol. Res. 7:5565-5572.
- Pinheiro EAA, Carvalho JM, Santos DCP (2013). Antibacterial activity of alkaloids produced by endophytic fungus *Aspergillus* sp. EJC08 isolated from medical plant *Bauhinia guianensis*. Nat. Prod. Res. 27:1633-1638.
- Pusz W, Płaškowska E, Yildirim I (2015). Fungi occurring on the plants of the genus *Amaranthus* L. Turk. J. Bot. 39:147-161.
- Queires LC, Crépin M, Vacherot F (2013). In vitro effects of polyphenols extracted from the aroeira plant (*Schinus terebinthifolius* raddi) on the growth of prostate cancer cells (LNCaP, PC-3 AND DU145). Braz. J. Med. Hum. Health 1:71-82.
- Robertson EAH, Cartwright RA, Oldschool M (1956). Phenolic substances of manufactured tea. I. fraction and paper chromatography of water-soluble substances. J. Sci. Food Agric. 8: 72-80.
- Rocha IV, Ferraz PD, Farias TG, Oliveira SR (2015). Resistance of bacteria isolated from equipment in an Intensive Care Unit. Acta Paul Enf, 5: 433-439.
- Rodrigues IMC, Souza FAPS, Ferreira FA. (2009). Estudo fitoquímico de *Senna alata* por duas metodologias. *Planta Daninha*, 27:507-513.
- Santiago IF, Alves TM, Rabello A et al. (2012). Leishmanicidal and antitumoral activities of endophytic fungi associated with the Antarctic angiosperms *Deschampsia antarctica* Desv. and *Colobanthis quitensis* (Kunth) Bartl. *Extremophiles* 16:95-103.
- Sharma OP, Darwra RK. (1991). Thin layer chromatographic separations of lantadenes, the pentacyclic triterpenoids from lantana (*Lantana camara*) plant. J. Chromat. 587:351-354.
- Shen S, Ding R, Zhou Y et al. (2014). Immunomodulatory Effects of Polysaccharide from Marine Fungus *Phoma* sp. YS4108 on T Cells and Dendritic Cells. *Mediators Inflamm.* 13p.
- Shen X, Zheng D, Gao J (2012). Isolation and evaluation of endophytic fungi with antimicrobial ability from *Phyllostachys edulis*. *Bangladesh J. Pharmacol.* 7:249-257.
- Shrestha P, Szaro TM, Bruns TD (2011). Systematic Search for Cultivable Fungi That Best Deconstruct Cell Walls of *Miscanthus* and Sugarcane in the Field. *Appl. Environ. Microbiol.* 77:5490-5504.
- Shukla S, Shukla H, Pandey AK. (2014). Screening of some phytopathogenic fungi for their antimicrobial potential. *W J Pharm. Pharmaceut Sci*, 3:2478-2494.
- Silva AB, Silva T, Franco ES et al (2010). Antibacterial activity, chemical composition, and cytotoxicity of leaf's essential oil from Brazilian pepper tree (*Schinus terebinthifolius*, Raddi). *Braz. J. Microbiol.* 41:158-163.
- Siqueira VM, Conti R, Araújo JM (2011). Endophytic fungi from the medicinal plant *Lippia sidis* Cham. and their antimicrobial activity. *Symbiosis*, 53:89-95.
- Strobel G, Sanjay KS, Riyaz-Ul-Hassan S (2011). An endophytic/pathogenic *Phoma* sp. from creosote bush producing biologically active volatile compounds having fuel potential. *FEMS Microbiol Lett*, 320:87-94.
- Tayung K, Sarkar M, Baruah P (2012). Endophytic fungi occurring in *Ipomoea carnea* tissues and their antimicrobial potentials. *Braz. Arch. Biol. Technol.* 55:653-660.
- Wagner H, Bladt S (1996). *Plant drug analysis – A thin layer chromatography atlas*. Munich, Germany: Springer.
- Wang Y, Dai C-C. (2011). Endophytes: a potential resource for biosynthesis, biotransformation, and biodegradation. *Ann Microbiol*, 61:207-215.
- White TJ, Bruns T, Lee S (1990). Amplification and direct sequencing of fungal RNA genes for phylogenetics. In: *PCR Protocols. A Guide to Methods and Applications*. Innis MA, Gelfand DH, Sninsky JJ, White TJ (eds). San Diego: Academic, pp.315-322.
- Woodford N, Livermore DM (2009). Infections caused by Gram-positive bacteria: a review of the global challenge. *J. Infect.* 59:S4-S16.
- Zhang H, Xiong Y, Zhao H et al. (2013a). An antimicrobial compound from the endophytic fungus *Phoma* sp. isolated from the medicinal plant *Taraxacum mongolicum*. *J. Taiwan Inst. Chem. Eng.* 44:177-181.
- Zhang X, Ding R, Zhou Y et al. (2013b). Toll-Like Receptor 2 and Toll-Like Receptor 4-Dependent Activation of B Cells by a Polysaccharide from Marine Fungus *Phoma* sp. YS4108. *PLoS ONE* 3:e60781.

Full Length Research Paper

A very high frequency of hepatitis B and C virus infections during an active screening campaign in Abidjan

Kouassi-M'Bengue A.^{1,2*}, Ouattara Abdoulaye³, Allah-Kouadio Emile^{4,5}, Sevede Daouda¹,
Doumbia Moussa¹ and Dosso M.^{1,2}

¹National Hepatitis Viruses Reference Center, Pasteur Institute of Cote d'Ivoire 01 BP 490 Abidjan 01, Cote d'Ivoire.

²Department of Microbiology, Teaching and Research Unit of Medical Sciences, Félix Houphouët Boigny University Abidjan, Ivory Coast.

³Unit of Epidemiology, Department of Epidemiology and Clinical Research, Pasteur Institute of Cote d'Ivoire.

⁴National Hepatitis Infection Control Program, Ministry of Public Health BPV 13 Abidjan, Ivory Coast.

⁵Department of Medicine, Teaching and Research Unit of Medical Sciences, Félix Houphouët Boigny University Abidjan; Unit of Hepatology- CHU Cocody Abidjan, Ivory Coast.

Received 17 November 2016, Accepted 8 December, 2016

Viral hepatitis is a serious public health problem affecting billions of people globally. Limited information is available on this issue in Cote d'Ivoire. The objectives of this study were to determine the prevalence and factors associated with hepatitis B virus (HBV) and hepatitis C virus (HCV) during an active screening campaign in Abidjan. A cross-sectional study was conducted at Pasteur Institute of Cote d'Ivoire from July 2015 to February 2016. The ethical clearance for this study was obtained from the National Ethical and Research Committee. An informed written consent was obtained from the participants of the study and administered a questionnaire related to the socio demographical information and risk factors of a possible route transmission HBV and hepatitis C virus (HCV). Blood samples were collected for the detection of HBS Ag, Ab-HBc IgG and Ab-HCV. Serological analyses were performed by Cobase 601 (Roche^R). Data were analyzed by R software. A total of 1801 patients were recruited; among them 138 children (7.7%) aged from 0 to 15 years and 1663 adults (92.3%). The sex ratio was 1.2 (964/837). The overall prevalence of HBsAg was 30.9% (557/1801) and 41.1% (702/1708) for Ab-HBc IgG. About HCV, the overall prevalence rate was 5.3% (95/1687), none of the children was HCV positive. The co-infection HBV/HCV rate was 0.95% (16/1687). HCV was associated with age and sexual risk behaviors. HBV was associated with gender, youth, sexual risk behaviors, and scarification. Our findings revealed a high prevalence of HBV. The measures to reduce the disease and its load transmission must be strengthened.

Key words: Hepatitis B virus, hepatitis C virus, seroprevalence, risk factors, Cote d'Ivoire.

INTRODUCTION

According to the World Health Organization, more than 240 million people are infected with the hepatitis B virus (HBV) worldwide, and the majority lives in developing countries (WHO, 2013), such as Côte d'Ivoire. Every year, there are more than 600000 deaths due to the

complications related to the infection. HBV's association with liver diseases, such as the primary liver carcinoma and cirrhosis, is clearly established (Hammad and Zaghloul, 2009). HBV and HCV are easily transmissible through sexual, parenteral, and vertical routes (Pozzetto

and O. Garraud, 2011). Several behavioral, environmental, and cultural factors may also be responsible for these infections (Kramvis and Kew, 2007). In Africa, after the vertical and sexual transmissions, HBV and HCV infections are due to cultural practices (levirate, sorority, sexual rituals, scarification, piercing, and tattoos) or medical surgeries (Simpore et al., 2007). Several authors have reported different prevalence of HBV and HCV among sub-populations in Cote d'Ivoire with estimates which vary according to the studied population and methods that are used. In fact, the authors reported that 12.1% HIV co-infected children (Rouet et al., 2008), 3.2% children without HIV; 7.53% blood donors; and 17.6% hemodialysis patients were seropositive for HBs Ag (Kouassi-M'Bengue et al., 2010, 2012, 2013). For HCV infection, the authors reported that 2.5% among blood donors (Kouassi-M'Bengue et al., 2012) were seropositive. However, there is no reliable national survey of HBV and HCV exposure in the population is most likely to benefit from the early detection, surveillance and treatment. Given this gap in our understanding of HBV and HCV in Cote d'Ivoire, we launched an active screening campaign in Abidjan in collaboration with the National Hepatitis Infection Control Programs and the support of a private partner.

The aim of this study was to determine the prevalence and factors associated with HBV and HCV in Abidjan (Cote d'Ivoire).

MATERIALS AND METHODS

A partnership of five years (2015-2020) was signed between Cote d'Ivoire government and an international pharmaceutical laboratory. As a result of this contract, Pasteur Institute of Côte d'Ivoire (IPCI), hosting the National Reference Center (CNR) of viral hepatitis, has been designated as a technical implementation partner. A national awareness campaign was launched through the media to get a larger sensitization of the whole population.

A cross-sectional study was conducted at IPCI from July 2015 to February 2016. Voluntary patients or subjects addressed by healthcare staff to IPCI were enrolled in the study. The subjects responded to a range of questions such as socio demographic information, such as age, marital status, profession, educational background (literacy, schooling), knowledge about hepatitis B and C virus, site of living; other data on the risk factors of the possible route of HBV and HCV transmission like the sharing of toothbrush and cutting objects, sexual risk behaviors, history of blood transfusion, tattooing or piercing body, scarification, and the history of HBV vaccination was obtained. Blood samples were collected for the detection of HBS Ag, Ab-HBc IgG and Ab-HCV. Serological analyses were performed by Cobas e 601 (Roche[®]).

Ethical issues

The ethical clearance for this study was obtained from the National Ethical and Research committee. We obtained an informed written

consent from all participants of the study. Biological parents or relatives have given their consent for infants.

Data analysis

Data were entered into Epi Info 3.5 and analyzed by R software. Qualitative variables were compared using the Fisher exact test and quantitative variables by analysis of variance. A percentage with 95% confidence interval (CI) was used to describe the prevalence. OR with 95% CI was calculated for each association. A p values less than 5% was considered as significant.

RESULTS

A total of 1801 patients were screened for HBsAg and 1687 for anti-HCV Ab. In all, the sex-ratio (M/F) = 1.15 and 1.16, respectively for HBs Ag detection and anti-HCV Ab. The overall mean age was 36 (\pm 15) years, ranging from 2 months to 85 years.

The overall prevalence of HBsAg was 30.9% (557 positive) in 1801 subjects. We noted that 41.1% (702/1708) subjects had positive Ab-HBc IgG without any detection of HBS Ag.

HBsAg prevalence in <20, 20-29, 30-39, 40-49 and >50 age groups was 13.4, 39.8, 37.4, 31.1 and 18.1%, respectively, as well as it showed an increase with age from 13.4% in <20 age group and 39.8% in 20-29 group, respectively, then a decrease from 37.4% in 30-39 age group to 18.1% in the older group >50 years old (Figure 1A). When the overall prevalence of HBsAg marker was analyzed according to gender, the prevalence was 36.7 and 24.3% in male and female subjects, respectively. The difference was found to be statistically significant (OR = 1.8, P < 0.01) (Figure 1B).

The seroprevalence of anti-HCV Ab was found to be 5.6%, corresponding to 95 seropositive subjects out of 1687 participants. The prevalence of anti-HCV in <20, 20-29, 30-39, 40-49 and >50 age groups was 1.2, 3.3, 2.9, 3.3 and 18%, respectively. For these subjects, the anti-HCV prevalence increased with age (Figure 2A).

The overall prevalence of anti-HCV in 781 female subjects was 5.4% (42 positive), which was lower than that in the 906 male subjects, 5.8% (53 positive). The difference was found not to be statistically significant (P=0.67) (Figure 2B).

Regarding the seropositivity of HBV and HCV dual carriage, the presence of both HBsAg and anti-HCV antibodies was noted in 16 cases out of 1687 tested subjects (0.9%). These 16 patients were male in 87.5% and female in 12.5%. The proportion of co-infection was the same among the 20-29 and 30-39 years (31.3%), for 40-49 and >50 age group it was respectively 12.5 and 25%.

*Corresponding author. E-mail: alphonsinembengue@pasteur.ci. Tel: 00225 05856242.

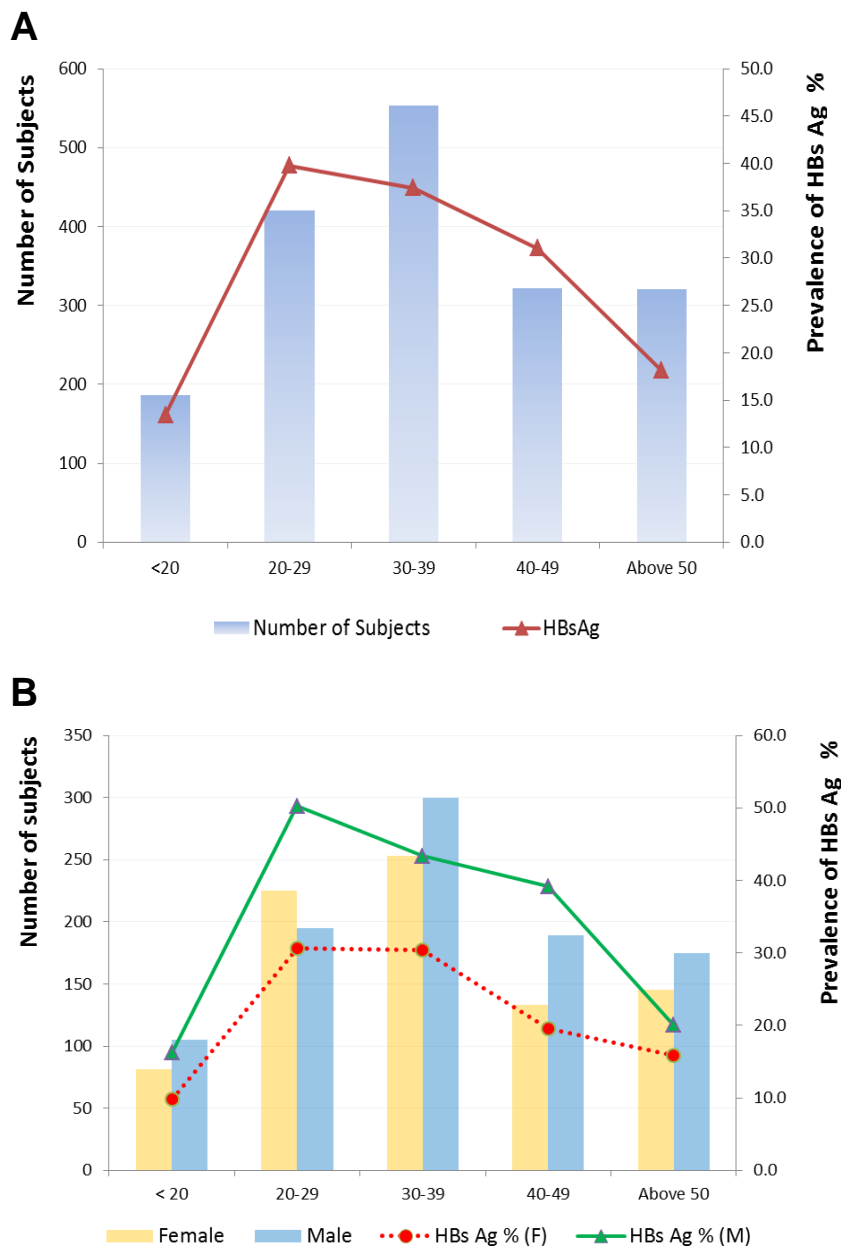


Figure 1. Serological features of HBsAg prevalence in Abidjan. Bars represent the number of subjects in each group indicated by left vertical axis, and the points represent the prevalence in each group indicated by right vertical axis. The horizontal axis represents the age ranges. **A**, Overall characterization of HBsAg by age group. **B**, Overall characterization of HBsAg by gender, M, male, F, female.

Table 1 lists the frequency of potential risk factors reported by groups (positive and negative) HBs Ag. On the subject of HBV carriage, HBsAg positivity was associated with male gender (OR= 1.8; $p < 0.01$), age (OR =2.9; $p < 0.01$), history of sexual risk behaviors (OR: 2.9; $p < 0.01$), family position (OR = 1.5; $p = 0.0005$) living in Abidjan (OR =1.5, $p=0.002$) and low socio economical level. It was defined by not salaried (OR=1.3; $p = 0.016$) and promiscuity (OR =1.3; $p=0.014$), scarification was also a risk factor (OR=1.6; $p<0.01$).

Conversely, there were no significant differences between HBV prevalence in people who benefited from a blood transfusion and those who have not (OR = 0.7; $p= 0.048$). That is the same thing for the people who were vaccinated against hepatitis B or not (OR=0.7; $p=0.013$). There was no significant difference between sharing toothbrush (OR = 1.2; $p =0.69$), sharing cutting objects (OR=0.96; $p=0.77$), body piercing (OR =1.3; $p=0.22$), tattooing (OR =1.2; $p=0.5$) a consumption of alcohol or drug (OR = 1.1 and 1.5; $p=0.39$ and 0.46). Literacy,

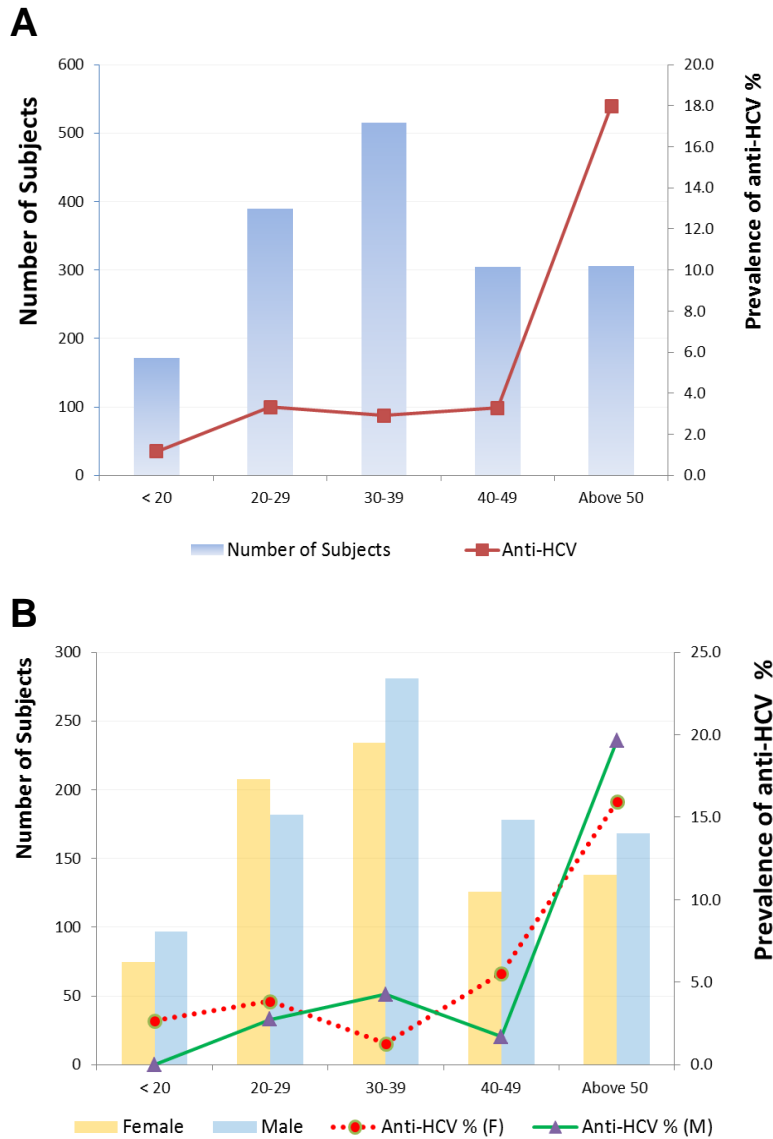


Figure 2. Serological features of anti-HCV prevalence in Abidjan. Bars represent the number of subjects in each group. Indicated by left vertical axis, and the points represent the prevalence in each group indicated by right vertical axis. The horizontal axis represents the age ranges. A, Overall characterization of anti-HCV by age group. B, Overall characterization of anti-HCV by gender, M, male, F, female.

schooling and knowledge about HBV are not associated with HBs Ag reactive with OR and p respectively 1.3, 0.12; 1.2, 0.008; and 0.96, 0.76.

Table 2 lists the frequency of potential risk factors reported by groups reactive or not with HCV-ab. Factors significantly associated with HCV infection were increasing age (OR =10.6; p<0.001) literacy (OR=3.5; p<0.001), schooling (OR= 2.5, p<0.001), living in Abidjan and family position. Conversely, no association was observed between HCV seropositivity and gender, tattooing, and sexual risk behaviors.

DISCUSSION

A cross-sectional study to assess the epidemiology of HCV and HBV prevalence among the general population living in Abidjan and other cities in Cote d'Ivoire was reported. To the best of our knowledge, this is the largest community-based epidemiologic study of HCV and HBV infections from Cote d'Ivoire during a national continual screening campaign. Most of the previous studies in this field were limited due to the short duration of the survey and scarce sample of populations (Assi et al., 2011). The

Table 1. Distribution of hepatitis B virus serological marker and principal risk factors in the study population; July 2015 to February 2016.

Risk Factor		HBV-Positive subjects		HBV-Negative subjects		OR (CI 95%)	p-value
		N	%	N	%		
Age	>=56	29	14.4	173	85.6	2.9 (1.9 – 4.6)	<0.001
	0-55	528	33	1071	70		
Sex	Female	203	24.3	634	75.7	1.8 (1.5 – 2.2)	<0.001
	Male	354	36.7	610	63.3		
Residence	Abidjan	440	29.4	1057	70.6	1.5 (1.2- 1.9)	0.002
	Out	117	38.5	187	61.5		
Family position	Couple	117	26	332	74	1.5 (1.2- 1.9)	0.0005
	Alone	412	35.2	759	64.8		
Salaried	Yes	327	30.4	749	69.6	1.3 (1.1- 1.6)	0.016
	No	213	36.3	374	63.7		
Promiscuity	Yes	145	36.3	254	63.7	1.3 (1.1- 1.7)	0.014
	No	389	29.9	914	70.1		
Sexual risk behaviors	Yes	436	34	846	66	1.7 (1.3- 2.2)	<0.001
	No	94	23.1	313	76.9		
Scarification	Yes	100	41	144	59	1.7 (1.3- 2.2)	<0.001
	No	419	29.4	1007	70.6		
Vaccination	Yes	91	26.5	252	73.5	0.7 (0.5-0.9)	0.013
	No	375	33.7	739	66.3		
Blood Transfusion	Yes	52	25.2	154	74.8	0.7 (0.5-0.99)	0.048
	No	474	32	1005	68		
Literacy	Yes	493	31.9	1052	68.1	1.3 (0.9-1.8)	0.12
	No	49	26.3	137	73.7		
Schooling	Basic	115	27.8	298	72.2	1.2 (0.9- 1.6)	0.08
	High	427	32.4	891	67.6		
Knowledge	Yes	369	32.3	775	67.7	0.96 (0.8- 1.2)	0.76
	No	133	33.1	269	66.9		
Drug	Yes	5	33.3	10	66.7	1.5 (0.5- 4.2)	0.46
	No	524	31.2	1158	68.8		
Tattoo	Yes	29	34.5	55	65.5	1.2 (0.7- 1.9)	0.5
	No	501	31	1114	69		
Body Piercing	Yes	33	37.1	56	62.9	1.3 (0.8- 2.1)	0.22
	No	496	30.9	1109	69.1		
Share toothbrush	Yes	11	34.4	21	65.6	1.2 (0.6- 2.4)	0.69
	No	516	31.1	1143	68.9		
Sharing of cutting objects	Yes	170	30.9	381	69.1	0.96 (0.8- 1.2)	0.77
	No	365	31.5	792	68.5		
Alcohol	Yes	159	32.8	326	67.2	1.1 (0.8- 1.4)	0.39
	No	360	30.6	815	69.4		

OR: Odds ratios, CI: confidence interval. Univariate analysis

Table 2. Distribution of hepatitis C virus serological marker and principal risk factors in the study population; July 2015 to February 2016

Risk factor		HCV-Positive subjects		HCV-Negative subjects		OR (CI 95%)	p-value
		N	%	N	%		
Age	>=56	49	25.3	145	74.7	10.8 (6.9 – 16.5)	<0.001
	0-55	46	3.1	1447	96.9		
Sex	Female	42	5.4	739	94.6	1.09 (0.7 – 1.7)	0.67
	Male	53	5.8	853	94.2		
Residence	Abidjan	69	4.9	1333	95.1	1.9 (1.2- 3.1)	0.005
	Out	26	9.1	259	90.9		
Family position	Couple	33	7.8	391	92.2	1.7 (1.0- 2.7)	0.03
	Alone	53	4.8	1042	95.2		
Salaried	Yes	63	6.3	936	93.7	1.1 (0.7- 1.7)	0.63
	No	32	5.7	530	94.3		
Promiscuity	Yes	24	6.3	358	93.7	1.3 (0.8- 2.0)	0.31
	No	60	5.0	1149	95.0		
Sexual risk behaviors	Yes	50	4.2	1148	95.8	0.5 (0.3- 0.8)	0.004
	No	30	7.9	350	92.1		
Scarification	Yes	12	5.2	221	94.8	1.00 (0.5- 1.9)	0.99
	No	68	5.1	1259	94.9		
Blood Transfusion	Yes	13	6.7	181	93.3	1.4 (0.7- 2.5)	0.32
	No	69	5.0	1313	95.0		
Literacy	Yes	61	4.2	138	95.8	3.5 (2.0- 5.9)	<0.001
	No	24	13.5	154	86.5		
Schooling	Basic	37	9.4	357	90.6	2.5 (1.6- 4.0)	<0.001
	High	48	3.9	1178	96.1		
Knowledge	Yes	47	4.5	1009	95.5	2.04 (1.29- 3.23)	0.002
	No	34	8.7	357	91.3		
Drug	Yes	1	6.7	14	93.3	1.3 (0.03- 8.7)	0.5
	No	82	5.2	1489	94.8		
Tattoo	Yes	1	1.3	77	98.7	0.2 (0.005- 1.3)	0.11
	No	82	5.4	1428	94.6		
Body Piercing	Yes	6	7.2	77	92.8	1.4 (0.5- 3.4)	0.26
	No	77	5.1	1423	94.9		
Share toothbrush	Yes	1	3.1	31	96.9	0.6 (0.01- 3.5)	0.48
	No	83	5.4	1465	94.6		
Alcohol	Yes	18	3.5	501	96.5	0.6 (0.3- 0.9)	0.03
	No	65	6.0	1013	94.0		

OR: Odds ratios, CI: confidence interval. Univariate analysis

descriptive epidemiologic data presented in this study can provide new insight into the contribution of HCV and HBV in the etiology of the liver disease in Cote d'Ivoire.

Data analysis revealed a very high prevalence (30.9%) of hepatitis B and confirmed a relative mean prevalence (5.3%) of hepatitis C. These results show a higher HBV and HCV prevalence than in the target groups as reported by other studies. So, Adoba et al. (2015) reported a prevalence of 14.5 and 0.5% for 200 barbers in Ghana; Coppola in a retrospective review study notified 0 to 15% HBV for immigrant coming from sub-Saharan Africa (Coppola et al., 2015); even in a retrospective study concerning 578 patients with chronic hepatitis conducted in Ethiopia, 22.3 and 3.6%, respectively for HBV and HCV were reported (Taye et al., 2015). However, regarding the rate of HCV, our results are lower than hemodialyzed and thalassemia patient with 33.5 and 37.1%, respectively (Malhotra et al., 2016; Ramadan et al., 2016).

Nevertheless, our rates are lower than the ones reported by Robert et al. (2016); in a population of HIV seropositive men which make sex with men with 49.4% for HBV and 10.4% for HCV. This high prevalence can be explained by the fact that it was the first year of national screening campaign and many people are interested. However, many patients are involved by medical prescription. A study realized in Kenya had recruited 389 patients with jaundice; HBs Ag seropositive was 50.6%, higher than our work (Missiani et al., 2016). Another study in Cameroon, concerning 97 hemodialyzed patients reported 6.2 and 20.6% seroprevalence for HBV and HCV (Halle et al., 2016).

This study also revealed a high rate of seropositivity to Ab-HBc-Ig G without any detection of HBs Ag in 41.1% cases. In this work, we did not research anti HBs, so our interpretation of these cases is a prior contact with HBV. The detection of the anti HBs Ab would have allowed us to decide between a passive immunization or the hypothesis of occult hepatitis. Concerning HBV and C co-infection, our prevalence was similar (0.9 vs. 0.2%) to that reported by Ojide in Nigeria (Ojide et al., 2015). The virological and molecular aspects of HBV/HCV coinfection are poorly comprehended. Although, liver disease activity and progression are generally more severe in the presence of double infection, an inverse relationship in the replicative. Levels of the two viruses exists, suggesting direct or indirect (that is, mediated by host immune responses) viral interference (Raimondo et al., 2008).

The analysis by gender has revealed that the seroprevalence of hepatitis B among males is significantly higher than that found in females. Our study is consistent with the results obtained by Deng et al. (2013) in 2013 in China (6.54% compared 3.87%) and Makuwa et al. (2009) in Gabon in 2009 (16.2% compared 9.9%); and no plausible explanation has been given for the higher rate in males in the general population but probably due to the

higher exposure to occupational HBV risk factors in men, or else females clear the HBV more efficiently as compared to males.

Age group of 16 to 55 years is the most affected (35%), compared to the older (14.4%). These results show that young people are the most affected by HBV, $p < 0.001$. These results are similar to those of Makuwa et al. (2009) who reported a prevalence of 22.22% among young men in the same age group in urban areas of Gabon. The low prevalence of individuals in the age group above 50 years of age could indicate that several people in this group might have died from cirrhosis or liver cancer due to the lack of medical care.

Regarding HCV, the higher prevalence was observed in advanced age groups; indeed, no anti-HCV seropositive case was found in 138 children younger than 16 years, nevertheless, the seropositivity increased progressively from adults (3.8%) to older persons (18%). The higher prevalence of HCV in older people could be attributed to a longer exposure to risk factors for HCV transmission; for instance iatrogenic transmission resulting from inadequately sterilized equipments, inappropriate use of supplies, etc. This risk factor is considered as the main risk associated with HCV infection in majority of the participants from the general population included in this study. This same risk is the primary cause for HCV transmission in many outbreaks documented in the United States and the European Union Health care (Centers for Disease Control and Prevention, 2011; Rantala, 2008).

This study also reports an HBV carriage in 12.3% of children under 16 years; a significant difference was observed between younger (0-5 years) and older children (6-15 years), (7.3% vs. 14.4%) $p = 0.25$. At this stage, the study cannot demonstrate the evidence of a vertical transmission or a horizontal infection. In fact, some traditional practices could explain the high prevalence of HBV in children, particularly, mothers using saliva to heal baby's wound. However, vertical transmission probably plays an important role, as in Cote d'Ivoire, no efficient action (HBV free screening during pregnancy, vaccination at birth and not at six weeks) is taken to fight against it.

The lack of significant difference in the prevalence of HBV among people when taking into account their history of blood transfusion can be explained by the improvement of blood safety in Cote d'Ivoire. In fact, HIV, hepatitis B and hepatitis C, and the bacterium *Treponema pallidum* subspecies *pallidum* are routinely detected in blood donations. However, the prevalence of 25.2% of HBV among transfused persons to 32% in non-transfused shows that the contamination by a residual risk of blood transfusion remains (Ouattara et al., 2006).

Conclusion

This study reports a high prevalence of HBV and HCV

carriage in Cote d'Ivoire. The results of our study showed that youth, gender, sexual risk behaviors, scarification, living out of Abidjan in one hand and in the other hand old age, sexual risk behaviors, literacy, schooling are the common risk factors for transmission, respectively of HBV and HCV infection in the community. A better organization and increased awareness campaigns on HBV and HCV will reduce their prevalence. The measures to reduce the disease and its load transmission must be strengthened.

Conflict of interests

The authors have not declared any conflict of interests.

ACKNOWLEDGEMENTS

The authors wish to acknowledge ROCHE WEST AFRICA for its support. The authors also gratefully acknowledge the study participants, whose availability and effort have enabled the achievement of this study.

REFERENCES

- Adoba Prince, Stephen Kyei Boadu, Hope Agbodzakey, Daniel Somuah, Richard Kobina Dadzie Ephraim, Enoch Anto Odame (2015). High prevalence of hepatitis B and poor knowledge on hepatitis B and C viral infections among barbers: A cross-sectional study of the Obuasi municipality, Ghana. *BMC Public Health* 15(1041):1-7
- Assi C, Ouattara A, Allah Kouadio E, Diakite M, Koné S, Lohoues Kouacou MJ, Camara BM (2011). Vaccination coverage against hepatitis B and prevalence of HBsAg: a cross-sectional study involving 592 persons attending public screening in Abidjan. *Clin. Res. Hepatol. Gastroenterol.* 35(6-7):506-7
- Centers for Disease Control and Prevention (CDC) (2011). Hepatitis C virus infection among adolescents and young adults: Massachusetts, 2002–2009. *MMWR Morb Mortal Wkly Rep*, 60(17):537-541.
- Coppola Nicola, Loredana Alessio, Mariantonietta Pisaturo, Margherita Macera, Caterina Sagnelli, Rosa Zampino, Evangelista Sagnelli (2015) -Hepatitis B virus infection in immigrant populations *World J Hepatol* 7(30):2955-2961
- Deng QJ, Pan YQ, Wang CY (2013). Prevalence and Risk Factors for hepatitis B in Hua County, Henan Province, Beijing Da Xue Xue Bao 45:965-970.
- Halle Marie Patrice, Simeon-Pierre Choukem, Francois Folefack Kaze, Gloria Ashuntantang, Vanessa Tchamago, Yannick Mboue-Djicka, Elvis Temfack, Henry N Luma, (2016). Hepatitis B, Hepatitis C, and Human Immune deficiency Virus Seroconversion Positivity Rates and Their Potential Risk Factors Among Patients on Maintenance Hemodialysis in Cameroon. *Iranian J. Kidney Dis.* 10(5):304-309
- Hammad AM, Zaghloul MHED (2009) "Hepatitis G virus infection in Egyptian children with chronic renal failure (single centre study) *Ann. Clin. Microbiol. Antimicrob.* 8:36.
- Kouassi-M'bengue A, Nassirou F, Doumbia M, Amorissani Folquet M (2010). Prévalence de l'hépatite virale B chez les enfants de moins de 5 ans en Côte d'Ivoire. *Revue Bio Africa*, (8):27-33.
- Kouassi-M'bengue Alphonsine, Siransy Bogui Liliane, Legbedji K Antoinette, Dembele Ba Mory, Doumbia Moussa, Konaté Séidou (2012). Abissé Agba Sébastien Séroprévalence des hépatites virales B et C, et de l'infection à VIH chez les donneurs de sang en Côte d'Ivoire en 2010, *Rev. Bio. Afr.* 10:40-46.
- Kouassi-M'Bengue A, Cissé B, Yao Hubert, Ouattara D, Doumbia M, Sevede D, Gnonsiahe (2013). Fréquence de l'hépatite virale B et du VIH chez les hémodialysés à Abidjan en 2010. *Revue Bio-Africa* 11: 43-48.
- Kouassi-M'Bengue A, Cissé B, Yao Hubert, Ouattara D, Doumbia M, Sevede D, (2013) Gnonsiahe Fréquence de l'hépatite virale B et du VIH chez les hémodialysés à Abidjan en 2010. *Rev. Bio-Afr.* 11:43-48
- Kramvis A Kew MC (2007). Epidemiology of hepatitis B virus in Africa, its genotypes and clinical associations of genotypes. *Hepatology Res.* 37(1):S9-S19
- Makuwa M, Mintsang-Ndong A, Souqui`ere S, Nkogh'e D, Leroy ME, Kazanji M (2009). Prevalence and molecular diversity of hepatitis B virus and hepatitis delta virus in urban and rural populations in northern Gabon in Central Africa. *J. Clin. Microbiol.* 47(7):2265-2268
- Malhotra R, Soin D, Grover P, Galhotra S, Khutan H, Kaur N (2016). Hepatitis B virus and hepatitis C virus co-infection in hemodialysis patients: A retrospective study from a tertiary care hospital of North India. *J. Nat. Sci. Biol. Med.* 7:72-74.
- Missiani Ochwoto, James H. Kimotho, Julius Oyugi, Fredrick Okoth, Henry Kioko, Simeon Mining, Nancy LM Budambula, Elizabeth Giles, Anton Andonov, Elijah Songok and Carla Osiowy (2016). Hepatitis B infection is highly prevalent among patients presenting with jaundice in Kenya *BMC Infectious Diseases* 16(1):1-14.
- Ojide CK, El Kalu E Ogbaini-Emevon VU Nwadike (2015). Co-infections of hepatitis B and C with human immunodeficiency virus among adult patients attending human immunodeficiency virus out patients clinic in Benin City, Nigeria. *Nigerian J. Clin. Pract.* 18(4):516-521
- Ouattara H, Siransy-Bogui L, Fretz C, Diane KM, Konate S, Koidio A, Minga KA, Hyda J, Koffi-Abe N, Offoumou AM, Abissey S (2006). Residual risk of HIV, HVB and HCV transmission by blood transfusion between 2002 and 2004 at the Abidjan National Blood Transfusion Center. *Transfusion Clin. Biol.* 13(4):242-245.
- Pozzetto B, Garraud O (2011). "Emergent viral threats in blood transfusion," *Transfusion Clinique et Biologique* 18(2):174–183,
- Raimondo G, Saitta C (2008). Treatment of the hepatitis B virus and hepatitis C virus co-infection: still a challenge for the hepatologist. *J Hepatol.* 49:677-679.
- Ramadan A. Mahmoud, Abdel-Azeem M. El-Mazary, Ashraf Khodeary (2016). Seroprevalence of Hepatitis C, Hepatitis B, Cytomegalovirus, and Human Immunodeficiency Viruses in Multi transfused Thalassemic Children in Upper Egypt *Advances in Hematology. Article ID 9032627*, 7 pp.
- Rantala M, van de Laar MJ (2008). Surveillance and epidemiology of hepatitis Band C in Europe - a review. *Euro Surveill*, 22:13-21
- Robert Remis S, Juan Liu, Mona Loutfy R, Wangari Tharao, Anuradha Rebbapragada, Sanja Huibner, Maya Kesler, Roberta Halpenny, Troy Grennan, Jason Brunetta, Graham Smith, Tatjana Reko, Rupert Kaul (2016). Prevalence of Sexually Transmitted Viral and Bacterial Infections in HIV-Positive and HIV Negative Men Who Have Sex with Men in Toronto *PLOS ONE* DOI:10.1371.
- Rouet F, Chaix ML, Inwoley A, Anaky MF, Fassinou P, Kpozehouen A, Rouzioux C, Blanche S, Msellati P (2008). Hepatitis B in African HIV-1–Coinfected Children. *Clin. Infect. Dis.* 46:361-366.
- Simpore J, Pietra V, Pignatelli S, Karou D, Nadembega WM, Ilboudo D, Ceccherini-Silberstein F, Ghilat-Avoid-Belem WN, Bellocchi MC, Saleri N, Sanou MJ, Ouedraogo CM, Nikiema JB, Colizzi V, Perno CP, Castelli F, Musumeci S (2007). Effective program against mother-to-child transmission of HIV at Saint Camille Medical Centre in Burkina Faso. *J. Med. Virol.* 79(7):873-879.
- Taye S, Abdulkerim A, Hussen M (2014). Prevalence of hepatitis B and C virus infections among patients with chronic hepatitis at Bereka Medical Center, Southeast Ethiopia: a retrospective study. *BMC Res. Notes* 7:272.
- WHO (2013). "Hepatitis B. Aide memoire", no. 204

Full Length Research Paper

Diversity and composition of methanotrophs in paddy soil as affected by different long-term fertilizer management from double-cropping paddy fields in Southern China

Haiming Tang^{1*}, Yilan Xu², Xiaoping Xiao¹, Jie Liu¹, Weiyan Li¹ and Jimin Sun¹

¹Hunan Soil and Fertilizer Institute, Changsha, 410125, China.

²College of Biological and Electromechanical Polytechnic, Changsha 410127, China.

Received 29 October, 2016; Accepted 8 December, 2016

Methane (CH₄) is the most important greenhouse gas, which was produced from paddy fields. The CH₄ production and emission were affected by methane-oxidizing bacteria (methanotrophs). Therefore, it is significant to investigate the effects of fertilizer applications on the change of soil methanotrophs, which affected CH₄ emission. The objective of this paper was to describe changes of CH₄ and diversity and composition of methanotrophs in paddy soil in relation to the application of crop residues, mineral fertilizer, and manure based on a long-term field experiment. In this study, static chamber-gas chromatography technique, real-time polymerase chain reaction (PCR) and Illumina high-throughput sequencing of the 16S rRNA gene, respectively, were used to analyze the CH₄ emissions from paddy fields, soil methanotrophs abundance and community diversity from May to October 2014 under five fertilization treatments: mineral fertilizer (MF), rice residue and mineral fertilizer (RF), low manure rate and mineral fertilizer (LOM), and high manure rate and mineral fertilizer (HOM), as compared to without fertilizer input (CK). The results indicated that CH₄ from fertilization treatments displayed different emission patterns during early and late rice growth period. HOM treatment had the highest CH₄ emissions during early and late rice growth period with 5.074 and 6.099 g m⁻², respectively. Some methanotrophs genera (*Methylosinus*, *Crenothrix*, *Methylocaldum*, *Methylomicrobium* and *Methylomonas*) were identified at the early and late rice main growth stages. The abundance and composition of soil methanotrophs were affected by long-term fertilization managements. The methanotrophs abundance was inhibited under MF treatment, while they were stimulated under RF, LOM and HOM treatments. The abundance and community composition of methanotrophs in paddy soil were affected by fertilizers of mineral, crop residues, and manure. It was concluded that application with organic and crop residues enhance the abundance and community composition of methanotrophs in double-cropping paddy fields in Southern China through a long-term fertilizer experiment.

Keywords: CH₄, long-term fertilization, methanotrophs diversity, methanotrophs composition, paddy field.

INTRODUCTION

Methane (CH₄) is the most greenhouse gas in the atmosphere and contributes approximately 18% to global

warming (IPCC, 2007). The atmospheric CH₄ concentration was affected by CH₄ production and oxidation. Paddy field is one of the major sources of CH₄, which annually emits 60 Tg CH₄ into the atmosphere (Lowe, 2006). Rice is the major food crops to feed people, especially in Asia (Krüger and Frenzel, 2003; Conrad et al., 2006). Therefore, some agronomic practices need to be strengthened and improved to obtain higher grain yield, such as fertilizer applications.

The CH₄ emission is affected by methanotrophs in the surface soil layer and rhizosphere, then it releases into the atmosphere. That is, CH₄ emission from paddy field was influenced by methanotrophs, which were gram-negative bacteria that utilize CH₄ as their sole source of carbon and energy (Lowe, 2006). According to the physiology, phylogeny, morphology and biochemistry, methanotrophs were classified into three main groups (type I, type II, and type X) (Hanson and Hanson, 1996). The growth and activity of methanotrophs was influenced by many factors, such as soil conditions, fertilizer application and vegetation cover (Hanson and Hanson, 1996; Zheng et al., 2008). And the fertilizer applications and rice plant are the important factors that affect growth and activity of methanotrophs.

At present, molecular approaches is widely used to assess the diversity and activity of methanotrophs, which used phylogenetic and functional gene probes to detect and analyze methanotrophs from samples (Murrell et al., 1998). In addition to the 16S rRNA gene, the presence and abundance of CH₄ oxidizers were also shown by functional genes of methanotrophs (Fjellbirkeland et al., 2001; Horz et al., 2001). For discriminating the methanotrophs, in earlier studies, different polymerase chain reaction (PCR) primers were designed to amplify 16S rRNA gene fragments of different groups CH₄ oxidizers (Henckel et al., 1999). High-throughput sequencing of the 16S rRNA gene was used to determine that the composition and diversity of microbial in response to soil tillage, different crop rotation and fertilizer applications. Soil methanotrophs community structure was changed after long-term fertilizer applications (Zheng et al., 2008). The relative data was used to measure abundance of operational taxonomic units (OTUs) and to calculate indices of richness and shannon of soil samples.

Our studies indicated that long-term fertilization managements could lead to significant changes in diversity of some soil microbe, enzyme activities, such as aerobic bacterial, actinomycete, fungus and β -glucosidase (Tang et al., 2014). Soil methanotrophs play an important role in carbon cycling in terrestrial ecosystems. The fertilizer application is the important factor that affects growth and activity of methanotrophs. A viable option was applied with manure and crop residue to changing CH₄

emissions, methanotrophs abundance and community composition in paddy field? Therefore, the objectives of this research were to: (1) CH₄ emissions from paddy fields respond to the long-term fertilization managements, and (2) long-term fertilizer managements lead to changes in methanotrophs abundance and community composition in paddy field. Therefore, the gas and soil samples were collected from a long-term fertilization experimental field and the CH₄ emissions from paddy fields, the methanotrophs abundance and community composition were studied using static chamber-gas chromatography technique, real-time PCR and Illumina high-throughput sequencing based on both 16S rRNA gene, respectively.

MATERIALS AND METHODS

Sites and cropping system

The experiment was established in 1986. It was located in Ning Xiang County (28°07' N, 112°18' E, and altitude 36 m) of Hunan Province, China. Under a continent monsoon climate, the annual mean precipitation was 1553 mm and potential evapotranspiration of 1354 mm. The monthly mean temperature was 17.2°C. Soil texture of the plough layer (0 to 20 cm) was silt clay loam with 13.71% sand and 57.73% silt. At the beginning of the study, the surface soil characteristics (0 to 20 cm) were as follows: soil organic carbon (SOC) 29.4 g kg⁻¹, total nitrogen 2.0 g kg⁻¹, available N 144.1 mg kg⁻¹, total phosphorous 0.59 g kg⁻¹, available P 12.87 mg kg⁻¹, total potassium 20.6 g kg⁻¹, and available potassium 33.0 mg kg⁻¹. There were three crops in a year, barley (*Hordeum vulgare* L.), early rice, and late rice (*Oryza sativa* L.). Barley was sown in the middle of November and harvested in early May of the following year. Early rice was then transplanted and harvested in the middle of July. The growth period of late rice lasted from late July to the end of October.

Experiment design

The experiment had five treatments: control (without fertilizer input, CK), mineral fertilizer (MF), rice residue and mineral fertilizer (RF), low manure rate and mineral fertilizer (LOM), and high manure rate and mineral fertilizer (HOM). The design ensured all fertilized treatments received equal N amount (the amount of N in mineral fertilizer plus that from rice residue or manure). The mineral fertilizers included urea, ordinary superphosphate, and potassium chloride. Details about the fertilizer management are listed in Table 1. Before rice transplanting seedling, manure and air-dried rice residue were incorporated into soil surface. The cultivation depth was about 20 cm. For early rice, and late rice, 70 and 60% of N was applied at seeding, and the remaining N was applied at top dressing stages. All the P₂O₅ and K₂O were applied at seeding stages. There were three replications and each plot size was 66.7 m². We referred to the data for the individual cropping periods from May to October, 2014.

1) For the RF treatment, rice straw return rate (air dry) was 2780 and 3600 kg ha⁻¹ for early and late rice. (2) For the LOM treatment, manure application rate (decomposed) was 2625.0 and 2670.0 kg

*Corresponding author. E-mail: tanghaiming66@163.com.

Table 1. Nutrient supply from rice straw, chicken manure and mineral fertilizer under different fertilizer treatments (kg ha⁻¹).

Treatments	Early rice			Late rice			Total		
	N	P ₂ O ₅	K ₂ O	N	P ₂ O ₅	K ₂ O	N	P ₂ O ₅	K ₂ O
CK	0+0 [*]	0+0	0+0	0+0	0+0	0+0	0	0	0
MF	142.5+0	54.0+0	63.0+0	157.5+0	43.2+0	81.0+0	300.0	97.2	144.0
RF	124.4+18.1	50.4+3.6	38.3+24.7	133.0+24.5	37.8+5.4	48.2+32.8	300.0	97.2	144.0
LOM	96.0+46.5	33.0+21.0	33.6+29.4	110.2+47.3	21.8+21.4	51.1+29.9	300.0	97.2	144.0
HOM	49.6+92.9	12.0+42.0	4.2+58.8	63.0+94.5	0.5+42.7	21.2+59.8	300.0	97.2	144.0

*Input from mineral fertilizer + input from organic fertilizer. The treatments are without fertilizer (CK), mineral fertilizer (MF), crop residue and mineral fertilizer (RF), low manure rate and mineral fertilizer (LOM), and high manure rate and mineral fertilizer (HOM).

ha⁻¹ for early and late rice. (3) For the HOM treatment, manure application rate (decomposed) was 5250.0 and 5340.0 kg ha⁻¹ for early and late rice. (4) The N, P₂O₅, and K₂O content of air-dry early rice straw was 0.65, 0.13, and 0.89%; N, P₂O₅, and K₂O content of air-dry late rice straw was 0.68, 0.15, and 0.91%, respectively; and N, P₂O₅, and K₂O content of decomposed chicken manure was 1.77, 0.80, and 1.12%, respectively.

Sample collection

Samples were collected in 2014. In each plot, soil samples in the rhizosphere soil were collected from the rice plants root at different rice growth stages. Three samples were collected from each plot. The samples were immediately frozen until further analyses could be performed.

CH₄ emissions from paddy fields were investigated using the static chamber-GC technique at 9:00 to 11:00 in the morning during rice growth period. From the second day after transplanting of early or late rice, gases sample were collected weekly.

Measurement of CH₄

The quantities of CH₄ emission were survey with a gas chromatograph (Agilent 7890A) equipped with flame ionization detector (FID). CH₄ was separated using 2 m stainless-steel column with an inner diameter of 2 mm 13×MS column (60/80 mesh), with FID at 200°C.

CH₄ fluxes were calculated with the following equation (Liebig et al., 2010):

$$F = \rho h [273 / (273+T)] dC / dt$$

where F is the CH₄ emission (mg m⁻² h⁻¹); T is the air temperature (°C) inside the chamber; ρ is the CH₄ density at standard state (0.714 kg m⁻³ for CH₄); h is the headspace height of the chamber (m); and dC/dt is the slope of the curve of gas concentration variation with time.

The total CH₄ emissions (g CH₄·m⁻²) were sequentially computed from the emissions between every 2 adjacent intervals of the measurements, based on a non-linear, least-squares method of analysis (Singh et al., 1996).

DNA Extraction and PCR

For each sample, DNA was isolated from 0.5 g of soil using the MoBio PowerSoil™ DNA Isolation Kit (Carlsbad, CA, USA). Extractions were executed according to the manufacturer's protocol. All genomic DNA concentration and purity were determined by

NanoDrop spectrophotometry (Thermo Scientific, Wilmington, DE, USA). PCR was performed at an initial denaturation temperature of 94°C for 3 min, followed by 20 cycles of 94°C for 45 s, 53°C for 30 s, and 65°C for 90 s. A final elongation step at 65°C was run for 10 min. PCR products were purified using the Qiagen™ PCR purification kit following the manufacturer's protocol with the exception of eluting in sterile water (Qiagen, Valencia, CA, USA) and quantified in Qubit 2.0 Fluorometer (Invitrogen, NY, USA).

Illumina high-throughput sequencing of 16S rRNA genes

Primers 515F and 806R (Caporaso et al., 2010) were used to target the V3–V4 region of the 16S rRNA gene with the addition of a barcoded sequence and the required Illumina adapters. Sequencing was performed on an Illumina (Illumina, Miseq–OE Biotech Company; Shanghai, China) with two paired-end read cycles of 300 bases each. Sequence analysis and OTUs identification was according to the methods of Giongo et al. (2010) and Fagen et al. (2012). Reads were trimmed to remove low quality bases and to remove the first 11 bases corresponding to the primer region by a script based on Trim2 (Huang et al., 2003), and then the reads were separated by barcode. Paired reads were assembled using FLASH to the reference greengenes (Reyon et al., 2012) 16S SSU rRNA database.

IFA analysis of methanotrophs bacteria

The methanotrophs genera were selected using the indirect immunofluorescence (IFA) method (Svetlana et al., 2007). Eight polyclonal antibodies specific for 8 methanotrophs genera, namely, *Methylosinus*, *Crenothrix*, *Methylobacter*, *Methylocaldum*, *Methylococcus*, *Methylomicrobium*, *Methylomonas*, and *Methylosarcina* were applied. The cross-reactivity and specificity of the antibodies were tested previously and found to be species-specific (Svetlana et al., 2007). The total number of methanotrophs was calculated as the sum of 8 pre-selected methanotrophs genera.

Diversity indices

To estimate bacterial diversity of each sample, the number of OTUs, richness and Shannon index were calculated using Mothur (Shannon, 1963). The phylogenetic distribution of OTUs was constructed by QIIME software.

Data analysis

The data were analyzed as a randomized complete block, using the

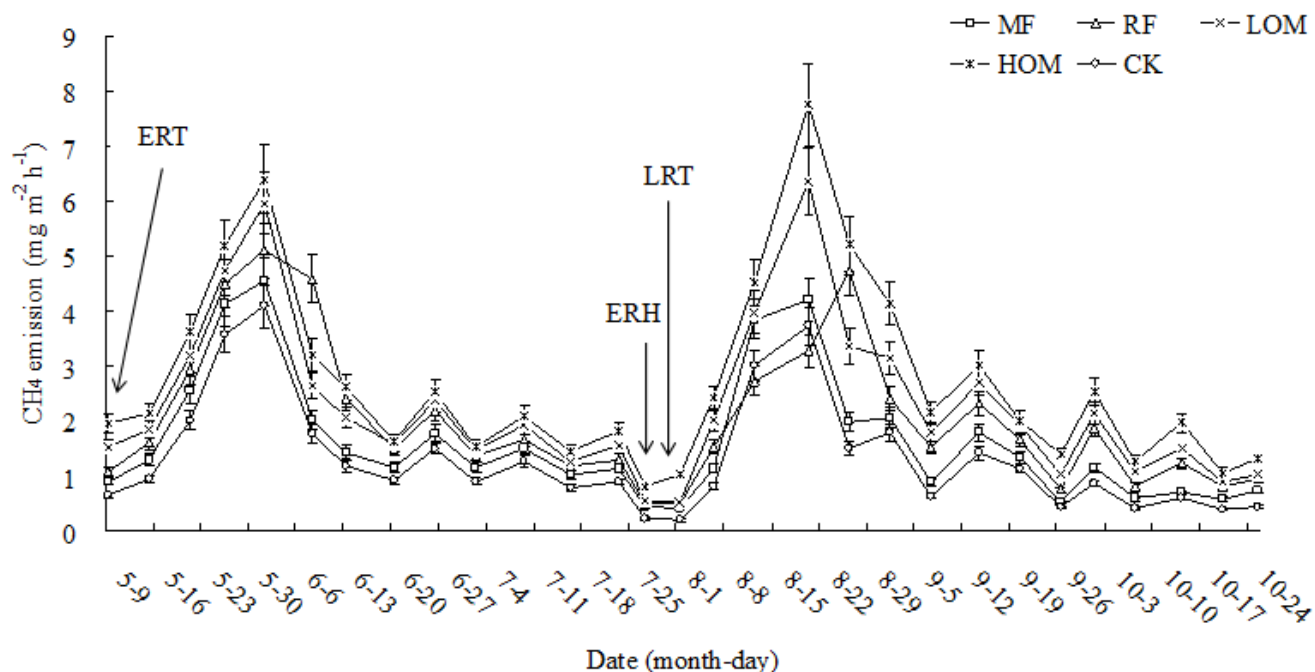


Figure 1. CH₄ emission from paddy fields affected by long-term fertilizer managements during early and late rice growth period. MF: Mineral fertilizer; RF: crop residue and mineral fertilizer; LOM: low manure rate and mineral fertilizer; HOM: high manure rate and mineral fertilizer; CK: without fertilizer. ERT: early rice transplanting; ERH: early rice harvesting; LRT: late rice transplanting. CH₄ emission rate is the mean of values measured within each treatment (n=3). Bars indicate standard deviation.

PROC ANOVA procedure of SAS (SAS Institute, 2003). Mean values were compared using the least significant difference (LSD) test and a probability value of 0.05 was considered to indicate statistical significance.

RESULTS

CH₄ emission

During early rice growing season, the CH₄ emission was low after early rice transplanting, but increased quickly until the peak appeared at 23 days after transplanting, and then declined to a low and stable level (Figure 1). The CH₄ emission were significantly different among treatments with the sequence of HOM>LOM>RF>MF>CK ($P<0.05$) during early rice growing season (Figure 1).

During late rice growing season, CH₄ emission mainly focused at tillering stage and the peak value of CH₄ emission was observed at 24 days after transplanting. Then, the emission rate decreased to a low level with relative stability. The sequence of treatments in CH₄ emission was HOM>LOM>RF>MF>CK (Figure 1).

During early rice growing season, the accumulate CH₄ emission of CK was significantly lower than MF, RF, LOM, and HOM ($P<0.05$) and the sequence with different treatments was HOM>LOM>RF>MF>CK (Table 2). The total CH₄ emissions from paddy fields during late rice growing season were 3.212 g m⁻² in MF, 3.961 g m⁻² in RF, 4.881 g m⁻² in LOM, 6.099 g m⁻² in HOM, and 2.548 g m⁻²

in CK. The order of treatments in total CH₄ emission was HOM>LOM>RF>MF>CK (Table 2). Meanwhile, HOM had larger total CH₄ emissions than other treatments during early and late rice growing season.

Operational taxonomic units of methanotrophs bacteria

At the early rice main growth stages, the root methanotrophs bacteria of rice plants from the LOM treatment had the highest number of OTUs (Figure 2). And the highest values of the OTUs in LOM with 76 at seedling stage (SS), the highest values of the OTUs in MF, RF, CK with 23, 41, and 47 at tillering stage (TS) among the different treatments. At the early rice main growth stages, the OTUs values were significantly different among treatments with the sequence of LOW>CK>RF>HOM>MF (Figure 2). At the late rice main growth stages, the highest values of the OTUs at SS in LOM and HOM with 53 and 40, and then dramatically declined to a low level. And the highest values of the OTUs were observed at TS in MF and RF with 39 and 62 after transplanting. Then, the OTUs values dramatically decreased to a low level.

Genetic diversity indices of methanotrophs

At early and late rice main growth stages, the root

Table 2. Effects of long-term fertilizer managements on CH₄ emission from paddy fields during whole growth period of early and late rice (g CH₄·m⁻²).

Treatments	Early rice	Late rice	Total
MF	3.470±0.147 ^c	3.212±0.176 ^d	6.682±0.323 ^d
RF	4.418±0.131 ^b	3.961±0.141 ^c	8.379±0.271 ^c
LOM	4.521±0.128 ^b	4.881±0.114 ^b	9.402±0.242 ^b
HOM	5.074±0.100 ^a	6.099±0.093 ^a	11.173±0.193 ^a
CK	2.886±0.083 ^d	2.548±0.074 ^e	5.434±0.157 ^e

MF: Mineral fertilizer; RF: crop residue and mineral fertilizer; LOM: low manure rate and mineral fertilizer; HOM: high manure rate and mineral fertilizer; CK: without fertilizer. Values are presented as mean ± SE (n = 3). Means in each column with different letters are significantly different at the $P < 0.05$ level.

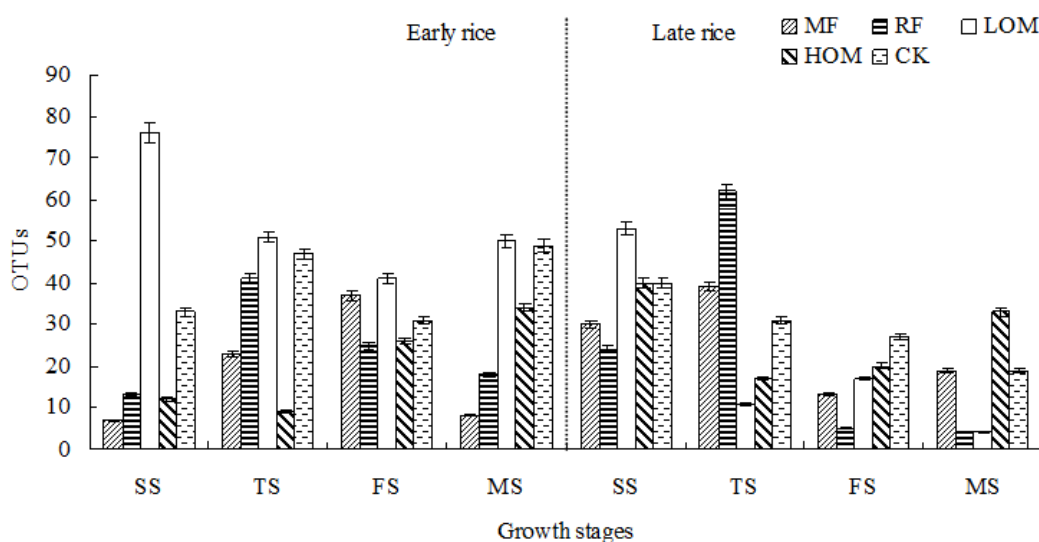


Figure 2. Operational taxonomic units of methanotrophs bacteria with different fertilizer treatments at rice main growth stages. SS: Seedling stage; TS: tillering stage; FS: full heading stage; MS: maturity stage. MF: mineral fertilizer; RF: crop residue and mineral fertilizer; LOM: low manure rate and mineral fertilizer; HOM: high manure rate and mineral fertilizer; CK: without fertilizer. Values are presented as mean ± SE (n = 3).

methanotrophs of rice plants from the paddy fields had the highest values of the richness indexes at TS and SS, respectively, and had the lowest values of the richness indexes at maturity stage (MS). Meanwhile, at early and late rice main growth stages, the values of the richness indexes were higher in HOM than in MF, RF, LOM and CK. The order of treatments in the values of the richness indexes was HOM>LOM>RF>CK>MF (Table 3).

At early and late rice main growth stages, the root methanotrophs of rice plants from the paddy fields had the highest values of the shannon indexes at TS and SS, respectively and had the lowest values of the shannon indexes at MS. Meanwhile, at the early and late rice main growth stages, the values of the shannon indexes were higher in HOM and LOM than in MF, RF and CK. The order of treatments in the values of the shannon indexes was HOM>LOM>RF>CK>MF (Table 3).

Structure of the methanotrophs bacteria community

It was indicated that some methanotrophs genera (*Methylosinus*, *Crenothrix*, *Methylobacter*, *Methylocaldum*, *Methylococcus*, *Methylomicrobium*, *Methylomonas* and *Methylosarcina*) were identified at main growth stages of early rice and late rice. It was shown that some methanotrophs genera were detected at some growth stages of early rice (Table 4). *Methylosinus*, *Crenothrix*, *Methylocaldum*, *Methylomicrobium* and *Methylomonas* were detected more often in all treatments at early rice main growth stages. *Methylobacter* were detected more often in LOM at the SS and MS. *Methylococcus* were detected more often in CK at main growth stages of early rice. *Methylosarcina* were detected more often in HOM and CK at early rice main growth stages.

Methylosinus, *Crenothrix*, *Methylomicrobium*,

Table 3. Genetic diversity indices of methanotrophs with different fertilizer treatments during rice main growth stages.

Rice	Treatment	Items							
		Richness index				Shannon index			
		SS	TS	FS	MS	SS	TS	FS	MS
Early rice	MF	22.4±0.65 ^b	25.4±0.73 ^b	19.7±0.57 ^{bc}	16.8±0.49 ^b	2.75±0.08 ^c	3.17±0.09 ^b	2.68±0.08 ^b	2.24±0.06 ^b
	RF	23.8±0.69 ^{ab}	26.8±0.77 ^{ab}	21.5±0.62 ^{ab}	17.8±0.51 ^{ab}	2.97±0.09 ^{ab}	3.45±0.10 ^{ab}	2.84±0.08 ^{ab}	2.42±0.07 ^{ab}
	LOM	24.3±0.70 ^{ab}	27.2±0.79 ^{ab}	21.9±0.63 ^{ab}	18.2±0.53 ^{ab}	3.06±0.09 ^a	3.57±0.10 ^a	2.92±0.08 ^{ab}	2.51±0.07 ^{ab}
	HOM	24.7±0.71 ^a	28.3±0.82 ^a	22.3±0.64 ^a	18.5±0.53 ^a	3.18±0.09 ^a	3.63±0.10 ^a	3.07±0.09 ^a	2.63±0.08 ^a
	CK	23.0±0.66 ^{ab}	26.0±0.75 ^{ab}	20.3±0.59 ^c	17.4±0.50 ^{ab}	2.85±0.08 ^{bc}	3.34±0.10 ^{ab}	2.75±0.08 ^b	2.38±0.07 ^b
Late rice	MF	23.4±0.68 ^a	20.8±0.60 ^b	18.7±0.54 ^b	15.1±0.44 ^b	2.82±0.08 ^c	2.73±0.08 ^c	2.67±0.08 ^c	2.21±0.06 ^c
	RF	24.7±0.71 ^a	22.3±0.64 ^{ab}	19.7±0.57 ^{ab}	15.8±0.46 ^{ab}	3.02±0.09 ^{ab}	2.97±0.09 ^{ab}	2.95±0.09 ^{ab}	2.47±0.07 ^{ab}
	LOM	25.2±0.73 ^a	22.8±0.66 ^{ab}	20.4±0.59 ^{ab}	16.2±0.47 ^{ab}	3.14±0.09 ^a	3.04±0.09 ^a	3.02±0.09 ^{ab}	2.58±0.07 ^a
	HOM	25.6±0.74 ^a	23.4±0.68 ^a	20.8±0.60 ^a	16.6±0.48 ^a	3.26±0.09 ^a	3.18±0.09 ^a	3.14±0.09 ^a	2.66±0.08 ^a
	CK	24.2±0.70 ^a	21.5±0.62 ^{ab}	19.4±0.56 ^{ab}	15.6±0.45 ^{ab}	2.95±0.09 ^{bc}	2.86±0.08 ^{bc}	2.84±0.08 ^{bc}	2.35±0.07 ^{bc}

SS: Seedling stage; TS: tillering stage; FS: full heading stage; MS: maturity stage. MF: mineral fertilizer; RF: crop residue and mineral fertilizer; LOM: low manure rate and mineral fertilizer; HOM: high manure rate and mineral fertilizer; CK: without fertilizer. Values are presented as mean ± SE (n = 3). Means in each column with different letters are significantly different at the $P < 0.05$ level.

Methylomonas and *Methylosarcina* were detected more often in all treatments at late rice main growth stages. *Methylocaldum* were detected more often in MF, RF, HOM and CK at main growth stages of late rice. *Methylococcus* were not detected in LOM and HOM at late rice main growth stages (Table 5).

Heatmap in the different fertilizer treatments during rice growth period

The 8 most abundant genera for five treatments at early rice main growth stages were analyzed and the differences were shown in the heatmap (Figure 3). The abundances of *Methylomonas* and *Crenothrix* were significantly higher in different treatments at main growth stages of early rice. *Methylosinus* were relatively more abundant in different treatments at SS. *Methylocaldum* were relatively more abundant in different treatments at SS and MS. On the other hand, *Methylobacter* and *Methylococcus* were not detected in different treatments at main growth stages of early rice. *Methylococcus* and *Methylobacter* were not detected in different treatments at full heading stage (FS).

At the late rice main growth stages, the relative abundances of *Methylomonas* were significantly higher in different treatments. *Methylosinus* were relatively more abundant in different treatments at SS. *Methylococcus* were relatively more abundant in different treatments at FS and MS. *Methylosarcina* were relatively more abundant in different treatments at MS. On the other hand, *Methylococcus* and *Methylocaldum* were not detected in different treatments at main growth stages of late rice. *Methylococcus* were less abundant in different treatments at SS and TS. *Crenothrix* were less abundant

in different treatments at MS (Figure 4).

Phylogenetic tree analysis of methanotrophs gene clones among different fertilizer treatments

At the early rice main growth stages, clustering analysis allowed the identification of OTUs responsible for the community shifts in different fertilization at genera level. The results showed that OTUs *Methylomonas* and *Crenothrix* were especially abundant in the soil samples. When phylogenetic trees were constructed using abundance belonging to each OTUs, *Methylocaldum* and *Methylosarcina* were grouped into a tight and distinct cluster, *Methylomonas* and *Crenothrix* were also grouped into a tight and distinct cluster. One-third of the clones in OTUs were grouped into a cluster including *Methylocaldum* (Figure 5).

At the late rice main growth stages, the results showed that OTUs *Methylomonas* and *Methylocaldum* were especially abundant in the soil samples. When phylogenetic trees were constructed using abundance belonging to each OTUs, *Crenothrix* and *Methylocaldum* were grouped into a tight and distinct cluster, *Methylomonas* and *Methylosarcina* were also grouped into a tight and distinct cluster. Quarter of the clones in OTUs was grouped into a cluster including *Methylocaldum* (Figure 5).

DISCUSSION

Effects of fertilizer applications on CH₄ emission

CH₄ emission is complex processes including production

Table 4. Genera of selected methanotrophs in different treatments at main growth stages of early rice.

Stage	Treatment	Methanotrophs							
		<i>Methylosinus</i>	<i>Crenothrix</i>	<i>Methylobacter</i>	<i>Methylocaldum</i>	<i>Methylococcus</i>	<i>Methylomicrobium</i>	<i>Methylomonas</i>	<i>Methylosarcina</i>
SS	MF	+	+	-	+	-	-	+	+
	RF	+	+	-	+	-	+	+	-
	LOM	+	+	+	+	-	+	+	-
	HOM	+	-	-	+	-	-	-	+
	CK	+	+	-	+	+	+	+	+
TS	MF	+	+	-	+	+	-	+	+
	RF	+	+	-	+	-	+	+	+
	LOM	+	+	-	+	-	+	+	+
	HOM	-	-	-	+	-	+	+	+
	CK	+	+	-	+	+	+	+	+
FS	MF	+	+	-	+	+	+	+	-
	RF	+	+	-	+	+	+	+	-
	LOM	+	+	-	-	-	+	+	-
	HOM	+	+	-	+	+	+	+	+
	CK	+	+	-	+	+	+	+	+
MS	MF	+	-	+	+	-	+	-	+
	RF	+	+	-	+	-	+	+	-
	LOM	+	+	+	+	+	+	+	+
	HOM	+	+	-	-	-	+	+	+
	CK	+	+	-	+	+	+	+	+

+ is present, it exist; - is absent, did not exist.

Table 5. Genera of selected methanotrophs in different treatments at main growth stages of late rice.

Stage	Treatment	Methanotrophs						
		<i>Methylosinus</i>	<i>Crenothrix</i>	<i>Methylocaldum</i>	<i>Methylococcus</i>	<i>Methylomicrobium</i>	<i>Methylomonas</i>	<i>Methylosarcina</i>
SS	MF	+	-	+	+	+	+	+
	RF	+	+	+	+	+	+	+
	LOM	+	+	-	-	+	+	+
	HOM	+	+	+	-	+	+	+
	CK	+	+	+	+	+	+	+

Table 5. Contd.

Stage	Treatment	Methanotrophs						
		<i>Methylosinus</i>	<i>Crenothrix</i>	<i>Methylocaldum</i>	<i>Methylococcus</i>	<i>Methylomicrobium</i>	<i>Methylomonas</i>	<i>Methylosarcina</i>
TS	RF	+	+	+	-	+	+	+
	LOM	+	+	-	-	+	+	-
	HOM	+	+	+	-	+	-	+
	CK	+	+	+	-	+	+	+
FS	MF	+	+	-	-	+	+	-
	RF	-	+	-	+	+	-	+
	LOM	+	+	+	-	+	+	+
	HOM	+	+	+	-	+	+	+
	CK	+	+	+	+	+	+	+
MS	MF	-	+	+	-	+	+	+
	RF	+	-	-	-	+	-	+
	LOM	+	-	-	-	-	+	+
	HOM	+	+	+	-	+	+	+
	CK	-	+	+	+	+	+	+

+ is present, it exist; - is absent, did not exist.

and oxidation. CH₄ production was regulated by vegetation type, fertilizer managements, soil temperature, soil moisture, root activity and many other factors (Wassmann et al., 2004; Kallenbach et al., 2010; Ma et al., 2008). In this study, the total CH₄ emission from paddy fields during rice growth period were much higher in HOM, LOM and RF as compared to CK (Figure 1 and Table 2), which was similar to the result by Wang et al. (2013). The reasons was (1) microbial activities were improved after returning rice residues, manure into the soil for that supplements of carbon source and energy for microbial activities to accelerate consumption of soil oxygen and decrease of soil redox potential (Eh); (2) methanogens became active for the large quantities of carbon source, which provided reactive substrate for CH₄ emission from paddy fields. Meanwhile, compared with the RF, LOM

and HOM, we also observed that MF decreased the CH₄ emission and resulted in a positive CH₄ emission from paddy fields. CH₄ emission was probably reduced because of excessive soil inorganic N level. Reduced root growth due to lower soil inorganic N level as a result of absence of N fertilization also probably reduced the methanotrophs activities, thereby resulting in lower CH₄ emission in MF. Several researchers (Bronson and Mosier, 1994; Powlson et al., 1997) have reported that N fertilization reduced CH₄ emission compared to no N fertilization. However, CH₄ emissions at early and late rice growth periods were decreased to a large extent after field drying, the reason may be (1) the methanogens activities were limited when the soil aeration was increased after field drying; and (2) the ability for transportation and emission of CH₄ was limiting,

which the rice plant physiological activities was decreased.

Effects of fertilizer applications on soil methanotrophs diversity

Organic and crop residues fertilization resulted in increasing methanotrophs bacteria diversity (Figure 2), showing that application of organic, crop residues significantly influenced methanotrophs bacteria composition, and mineral fertilization influenced methanotrophs bacteria composition to a lesser degree.

The response of methanotrophs bacteria community structures to agricultural managements was also represented by Nicol et al. (2003). In this study, it was shown that higher number of OTUs

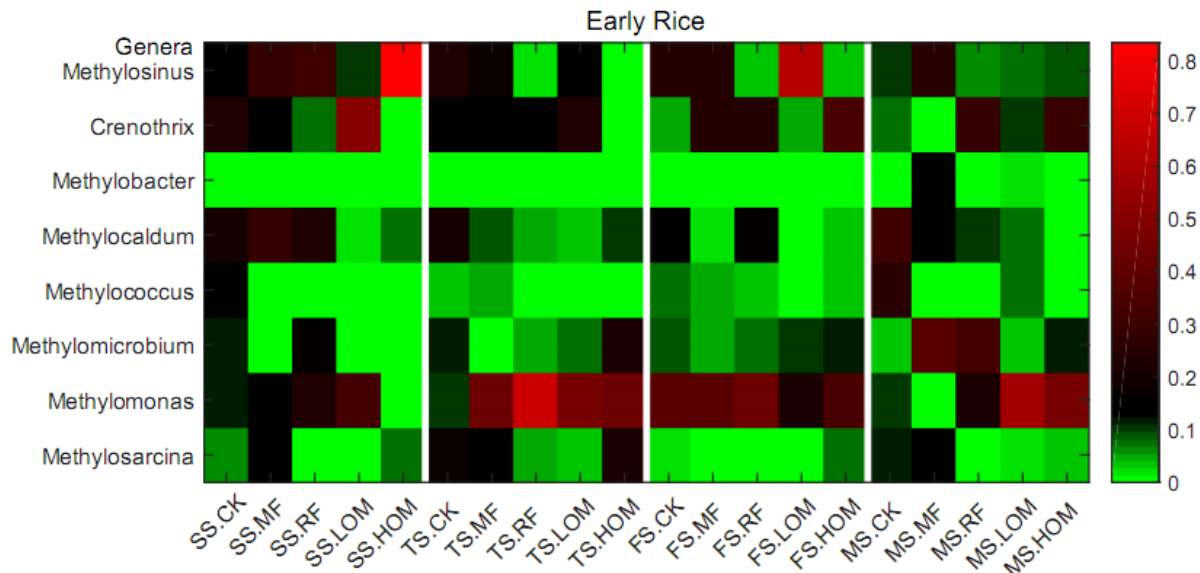


Figure 3. Heatmap illustrating the 8 most abundant methanotrophs genera in the different fertilizer treatments at early rice main growth stages. SS: seedling stage; TS: tillering stage; FS: full heading stage; MS: maturity stage. MF: mineral fertilizer; RF: crop residue and mineral fertilizer; LOM: low manure rate and mineral fertilizer; HOM: high manure rate and mineral fertilizer; CK: without fertilizer.

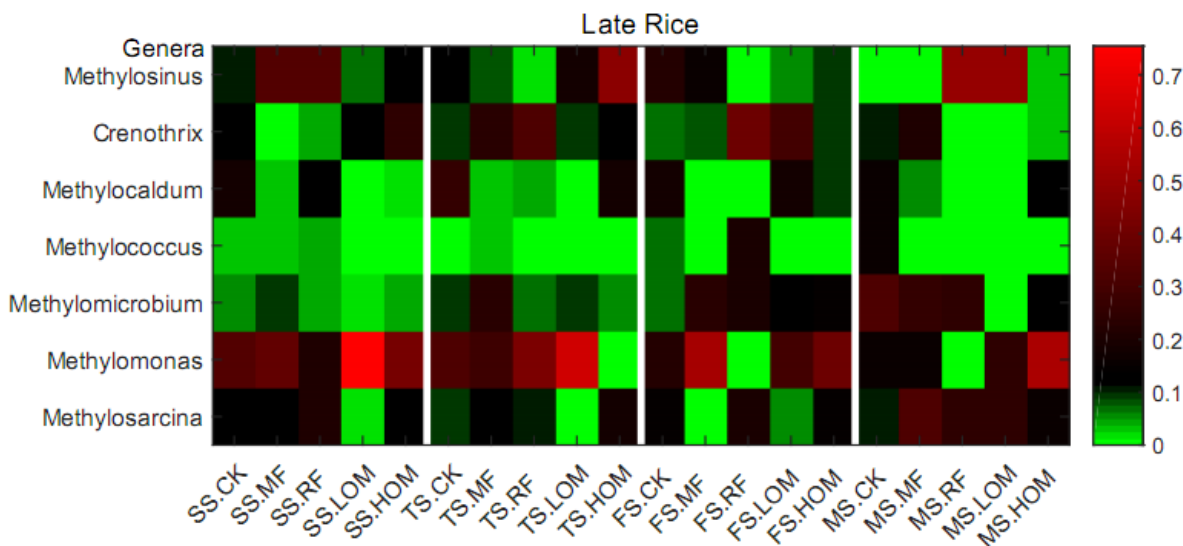


Figure 4. Heatmap illustrating the 7 most abundant methanotrophs genera in the different fertilizer treatments at late rice main growth stages. SS: seedling stage; TS: tillering stage; FS: full heading stage; MS: maturity stage. MF: mineral fertilizer; RF: crop residue and mineral fertilizer; LOM: low manure rate and mineral fertilizer; HOM: high manure rate and mineral fertilizer; CK: without fertilizer.

with organic, crop residues, than that of the mineral fertilizer. The relationship of methanotrophs bacteria and organic, crop residues, and mineral fertilizer in the experiment soil, proposes that organic crop residues is an important nutrient for the found taxa, which is relative to methanotrophs bacteria growth processes.

The methanotrophs microbial diversity was shown by

the richness index and shannon index which was larger in HOM, LOM, and RF treatments, which agree with Zheng et al. (2008), who found that soils under NPK and recycled crop residues had higher levels of methanotrophs diversity compared with the conventional fertilization. Meanwhile, shannon index and richness index was significantly higher when application with organic and crop

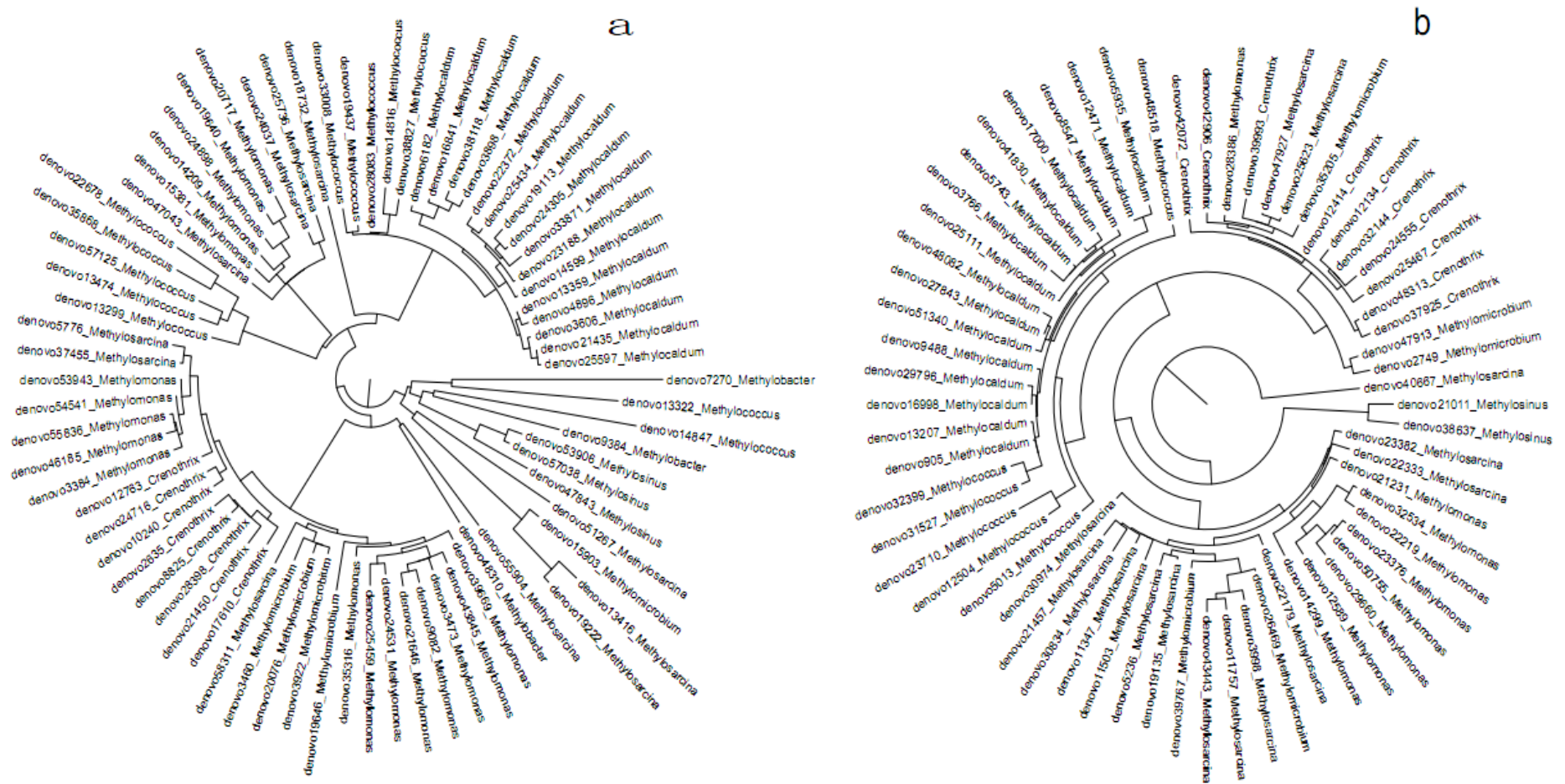


Figure 5. Phylogenetic distribution of OTUs of the clone libraries of 16S ribosomal RNA genes for soil samples under different treatments at early rice (a) and late rice (b) main growth stages.

residues, indicating that the application of organic and crop residues based on application of NPK, with higher rate of C:N, stimulates methanotrophs diversity on the soil. That is, rate of C:N in rice and organic straw substrates is higher than that of other mineral fertilizer (Garcia and Rice, 1994),

which provides more substrate to microbial explaining higher shannon index and richness index found in this work on plots where rice was the growing season. Meanwhile, it was key factors that root exudates influenced methanotrophs communities, for that they provide the carbon

source to soil microbial (Badri and Vivanco, 2009). Some studies indicated that roots may regulate the soil methanotrophs community in their rhizosphere, change the soil physical and chemical properties, and control the growth of competing plant species (Nardi et al., 2000), which may also affect the

methanotrophs diversity in rhizosphere.

Effects of fertilizer applications on soil methanotrophs community

According to the effects of different fertilizer treatments on methanotrophs abundance, five treatments (MF, RF, LOM, HOM and CK) were chosen to analyze the methanotrophs community structure. Some distinct differences in the diversity indices were found between the mineral fertilizer and application of organic; crop residues fertilizer treatments indicated that the soil methanotrophs community was changed with different fertilization managements. And the diversity pattern was shown in HOM and LOM treatments. In addition to the chemical NPK, the recycled application of crop residues and organic proposal were important to maintain methanogenic bacteria in paddy soils (Conrad and Klose, 2006). It is well-known that the soil CH₄ oxidation rate can be changed by different CH₄ concentrations, thus the growth, activities and community structure of methanotrophs were changed (Bender and Conrad, 1995). In conclusion, according to sequences and phylogenetic analysis, it was shown that soil methanotrophs community structure was changed by different fertilizer treatments.

Some study indicated that the genera of methanotrophs were different between paddy soils and forest soils (Mohanty et al., 2006). In this study, the 8 most abundant genera for five treatments at early rice main growth stages were shown in the heatmap (Figure 3), and the 7 most abundant genera for five treatments at late rice main growth stages were showed in the heatmap (Figure 4), indicated that composition significantly varied among the five estimated treatments (MF, RF, LOM, HOM and CK). Our results showed that methanotrophs were related to the genera of *Methylosinus*, *Crenothrix*, *Methylocaldum*, *Methylomicrobium* and *Methylomonas* with the most usual methane oxidizers in paddy soil with different fertilizer treatments, which were in agreement with an earlier study (Fjellbirkeland et al., 2001).

Conclusions

The results indicated that with the same nitrogen application rate, different organic-inorganic mixed fertilizer application, such as RF, LOM, and HOM, caused substantial CH₄ emissions during early and late rice growth period compared with those from the conventional MF treatment. Meanwhile, the abundance and composition of methanotrophs was affected by long-term fertilization managements. In the MF treatment, methanotrophs abundance was inhibited, compared to the RF, LOM and HOM treatments. The methanotrophs diversity of the HOM and LOM treatments was distinguished from RF, MF and CK. Furthermore, methanotrophs community composition was changed with

different treatments based on the sequences and phylogenetic analysis. And the higher ratio genera of *Methylosinus*, *Crenothrix*, *Methylomicrobium*, *Methylocaldum* and *Methylomonas* were found in the five treatments. In summary, the application of mineral fertilizer was an important factor that affected the abundance of methanotrophs and application of organic; crop residues enhance the abundance of methanotrophs in double-cropping paddy fields in Southern China. It is important to understand that the main effective factors for abundance and composition of methanotrophs, was linked to soil ecosystem processes and sustainable developing management of rice cultivation.

Conflict of Interests

The authors have not declared any conflict of interest.

ACKNOWLEDGEMENTS

This study was supported by the National Natural Science Foundation of China (No. 31571591, 31201178) and the Public Research Funds Projects of Agriculture, Ministry of Agriculture of the P.R. China (No. 201503123).

REFERENCES

- Badri DV, Vivanco JM (2009). Regulation and function of root exudates. *Plant Cell Environ.* 32(6):666-681.
- Bender M, Conrad R (1995). Effect of CH₄ concentrations and soil conditions on the induction of CH₄ oxidation activity. *Soil Biol. Biochem.* 27(12):1517-1527.
- Bronson KF, Mosier AR (1994). Suppression of methane oxidation in aerobic soil by nitrogen fertilizers, nitrification inhibitors, and urease inhibitors. *Biol. Fertil. Soils* 17(4):263-268.
- Caporaso JG, Lauber CL, Walters WA, Berg-Lyons D, Lozupone CA, Turnbaugh PJ, Fierer N, Knight R (2010). Global patterns of 16S rRNA diversity at a depth of millions of sequences per sample. *Proc. Natl. Acad. Sci. USA.* 108(1):4516-4522.
- Conrad R, Erkel C, Liesack W (2006). Rice Cluster I methanogens, an important group of Archaea producing greenhouse gas in soil. *Curr. Opin. Biotechnol.* 17(3):262-267.
- Conrad R, Klose M (2006). Dynamics of the methanogenic archaeal community in anoxic rice soil upon addition of straw. *Eur. J. Soil Sci.* 57(4):476-484.
- Fagen JR, Giongo A, Brown CT, Davis-Richardson A, Gano KA, Triplett EW (2012). Characterization of the relative abundance of the citrus pathogen *Ca. Liberibacter asiaticus* in the microbiome of its insect 525vecor, *Diaphorina citri*, using high throughput 16S rRNA sequencing. *Open Microbiol. J.* 6(1):29-33.
- Fjellbirkeland A, Torsvik V, Øvreås L (2001). Methanotrophic diversity in an agricultural soil as evaluated by denaturing gradient gel electrophoresis profiles of *pmoA*, *mxsA* and 16S rDNA sequences. *Antonie van Leeuwenhoek* 79(2):209-217.
- Garcia FO, Rice CW (1994). Microbial biomass dynamics in tall cereals prairie. *Soil Sci. Soc. Am. Pro.* 58(3):816-823.
- Giongo A, Crabb DB, Davis-Richardson AG, Chauhiac D, Mobberley JM, Gano K, Mukherjee N, Casella G, Roesch LF, Walts B, Riva A, King G, Triplett EW (2010). PANGEA: pipeline for analysis of next generation amplicons. *ISME J.* 4(7):852-861.
- Hanson HS, Hanson TE (1996). Methanotrophic bacteria. *Microbiol. Rev.* 60(2):439-471.

- Henckel T, Friedrich M, Conrad R (1999). Molecular analyses of the methane-oxidizing microbial community in rice field soil by targeting the genes of the 16S rRNA, particulate methane monooxygenase, and methanol dehydrogenase. *Appl. Environ. Microbiol.* 65(65):1980-1990.
- Horz HP, Yimga MT, Liesack W (2001). Detection of methanotroph diversity on roots of submerged rice plants by molecular retrieval of *pmoA*, *mmoX*, *mxαF*, and 16S rRNA and ribosomal DNA, including *pmoA*-based terminal restriction fragment length polymorphism profiling. *Appl. Environ. Microbiol.* 67(9):4177-4185.
- Huang X, Wang J, Aluru S, Yang SP, Hillier L (2003). PCAP: a whole genome assembly program. *Genome Res.* 13(9):2164-2170.
- IPCC-Intergovernment Panel on Climate Change (2007). *Climate Change 2007: the physical science basis. Working Group I contribution to the fourth assessment report of the intergovernmental panel on climate change.* Cambridge University Press, Cambridge.
- Kallenbach CM, Rolston DE, Horwath WR (2010). Cover cropping affects soil N₂O and CO₂ emissions differently depending on type of irrigation. *Agric. Ecosyst. Environ.* 137(3-4):251-260.
- Krüger M, Frenzel P (2003). Effects of N-fertilisation on CH₄ oxidation and production, and consequences for CH₄ emissions from microcosms and rice fields. *Glob. Change Biol.* 9(5):773-784.
- Liebig MA, Tanaka DL, Gross JR (2010). Fallow effects on soil carbon and greenhouse gas flux in central North Dakota. *Soil Sci. Soc. Am. J.* 74(2):358-365.
- Lowe DC (2006). Global change: a green source of surprise. *Nature* 439(7073):148-149.
- Ma J, Xu H, Yagi K, Cai ZC (2008). Methane emission from paddy soils as affected by wheat straw returning mode. *Plant Soil* 313(1):167-174.
- Mohanty SR, Bodelier PLE, Floris V, Conrad R (2006). Differential effects of nitrogenous fertilizers on methane-consuming microbes in rice field and forest soils. *Appl. Environ. Microbiol.* 72(2):1346-1354.
- Murrell JC, McDonald IR, Bourne DG (1998). Molecular methods for the study of methanotroph ecology. *FEMS Microbiol. Ecol.* 27(2):103-114.
- Nardi S, Concheri G, Pizzeghello D, Sturaro A, Rella R, Parvoli G (2000). Soil organic matter mobilization by root exudates. *Chemosphere* 41(5):653-658.
- Nicol GW, Glover LA, Prosser JI (2003). The impact of grassland management on archaeal community structure in upland pasture rhizosphere soil. *Environ. Microbiol.* 5(3):152-162.
- Powlson DS, Goulding KWT, Willison TW, Webster CP, Hutsch BW (1997). The effect of agriculture on methane oxidation in soil. *Nutr. Cycling Agroecosyst.* 49(1):59-70.
- Reyon D, Tsai SQ, Khayter C, Foden JA, Sander JD, Joung JK (2012). FLASH Assembly of TALENs enables high-throughput genome editing. *Nat. Biotechnol.* 30(5):460-465.
- Shannon CE (1963). *The mathematical theory of communication.* MD. Comput. 14:306-317.
- Singh JS, Singh S, Raghubanshi AS, Saranath S, Kashyap AK (1996). Methane flux from rice/wheat agroecosystem as affected by crop phenology, fertilization and water lever. *Plant Soil* 183(2):323-327.
- Svetlana B, Pascal B, Irina K, Valery G, Oswald VC (2007). Response of CH₄ oxidation and methanotrophic diversity to NH₄⁺ and CH₄ mixing ratios. *Biol. Fertil. Soils* 43(3):341-348.
- Tang HM, Xu YL, Sun JM, Xiao XP, Wang K, Li WY, Tang WG, Yang GL (2014). Soil enzyme activities and soil microbe population as influenced by long-term fertilizer management under an intensive cropping system. *J. Pure Appl. Microbiol.* 8(2):15-23.
- Wang JY, Chen ZZ, Ma YC, Sun LY, Xiong ZQ, Huang QW, Sheng QR (2013). Methane and nitrous oxide emissions as affected by organic-inorganic mixed fertilizer from a rice paddy in southeast China. *J. Soils Sediments* 13(8):1408-1417.
- Wassmann R, Neue HU, Ladha JK, Aulakh MS (2004). Mitigating greenhouse gas emissions from rice-wheat cropping systems in Asia. *Environ. Dev. Sustain.* 6(1):65-90.
- Zheng Y, Zhang LM, Zheng YM, Di HJ, He JZ (2008). Abundance and community composition of methanotrophs in a Chinese paddy soil under long-term fertilization practices. *J. Soils Sediments* 8(6):406-414.

African Journal of Microbiology Research

Related Journals Published by Academic Journals

- *African Journal of Biotechnology*
- *African Journal of Biochemistry Research*
- *Journal of Bacteriology Research*
- *Journal of Evolutionary Biology Research*
- *Journal of Yeast and Fungal Research*
- *Journal of Brewing and Distilling*

academicJournals